Research article

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Pholcid spiders from the Lower Guinean region of Central Africa: an overview, with descriptions of seven new species (Araneae, Pholcidae)

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Abstract. This paper summarizes current knowledge about Central African pholcids. Central Africa is here defined as the area between 10°N and 7°S and between 6°E and 18°E, including mainly the Lower Guinean subregion of the Guineo-Congolian center of endemism. This includes all of Gabon, Equatorial Guinea, São Tomé and Príncipe, most of Cameroon and Congo Republic, and parts of the neighboring countries. An annotated list of the 14 genera and 79 species recorded from this area is given, together with distribution maps and an identification key to genera. Seven species are newly described: *Anansus kamwai* sp. nov., *Leptopholcus gabonicus* sp. nov., *Ninetis faro* sp. nov., *Pholcus punu* sp. nov., *P. rawiriae* sp. nov., *Spermophora abibae* sp. nov., and *S. awalai* sp. nov. Additional new records are given for 16 previously described species, including 17 new country records. Distribution and diversity patterns are compared with data on West and East Africa. While West Africa contains a similar set of genera it is significantly less diverse than Central Africa. East Africa is taxonomically more distinct. It has similar levels of diversity as Central Africa, but appears to be less undersampled.

Keywords. Pholcidae, Central Africa, Lower Guinea, taxonomy, identification key

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Introduction

Central Africa is home to the world's second largest moist tropical forests, exceeded in area only by the Amazon Basin forests. In contrast to West and East Africa, where only about 10% of the original tropical moist forests remain (Naughton-Treves & Weber 2001; Norris *et al.* 2010), Central African forests may still extend over about half the original area (59.3% in 1992; Naughton-Treves & Weber 2001). While

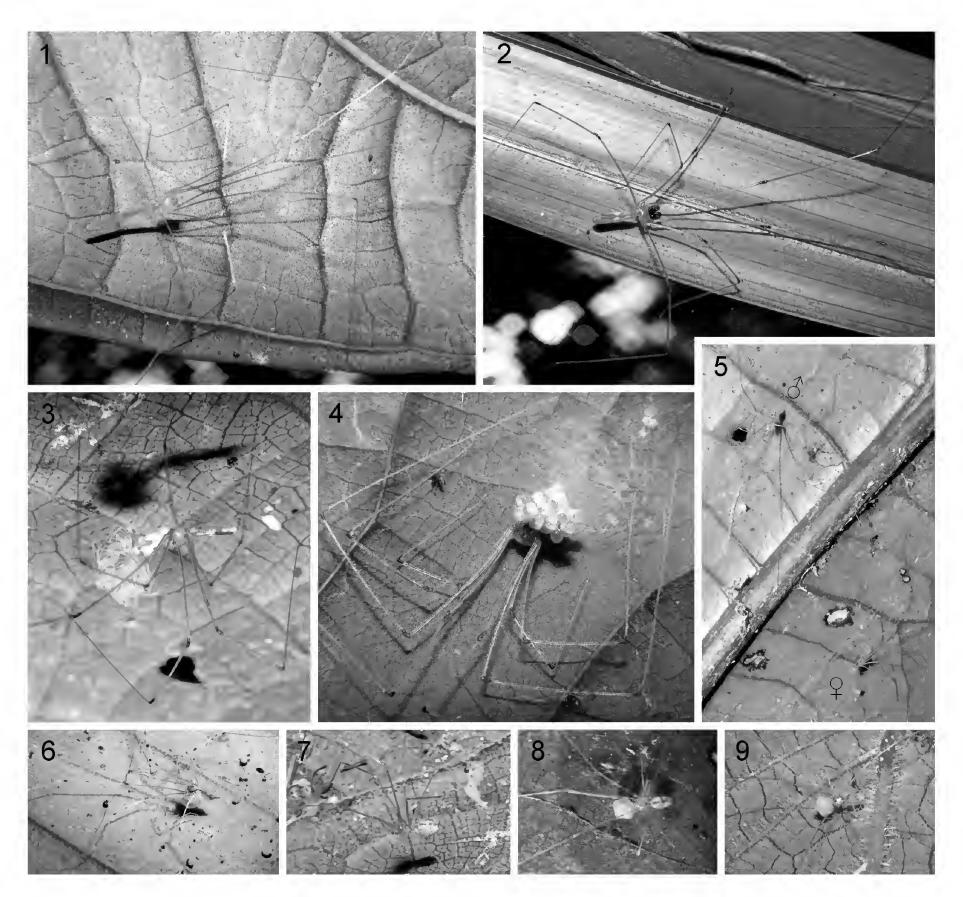
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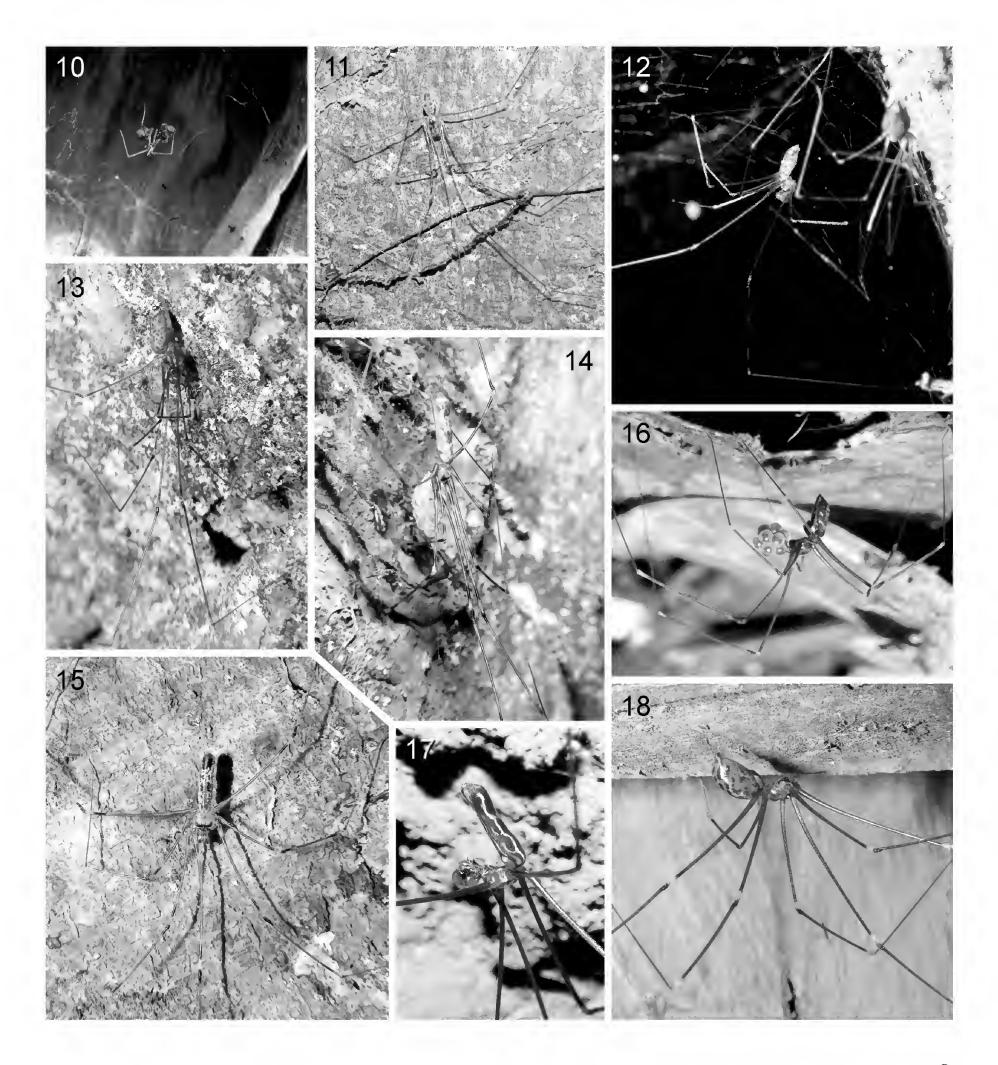
a high diversity of organisms otherwise concentrated in the tropics is thus expected for Central Africa, available data often fail to reflect this, probably for lack of exploration. Even relatively well-studied groups such as mammals, birds, and amphibians are poorly studied in parts of Africa, in particular in Central Africa (Dowsett-Lemaire & Dowsett 2001; Grubb 2001; Lawson & Klemens 2001), where cryptic diversity is probably more frequent than previously thought (Voelker *et al.* 2013).

Pholcid spiders are among the groups known to be most diverse in tropical and subtropical regions (Huber 2011a, 2011b), with a wide range of body shapes and colors related to different microhabitats (Figs 1-18). However, tropical Africa has long remained poorly explored (Huber 2005). No more than 18 currently valid species had been described from the Lower Guinean subregion of the Guineo-Congolian



Figs 1-9. Leaf-dwelling Central African pholcids. **1**. *Leptoholcus gabonicus* sp. nov., \Diamond from Mayebout, Gabon. **2-3**. *Pholcus baka* Huber, 2011, \Diamond from Sahoué (2) and \Diamond with juveniles from Mayebout (3), Gabon. **4**. *Smeringopus cylindrogaster* (Simon, 1907), \Diamond with eggsac from Loum, Cameroon. **5-6**. *Spermophora awalai* sp. nov., pair from Nyasoso (5) and a \Diamond with eggsac from Attuleh (6), Cameroon. **7**. *Spermophora akwamu* Huber & Kwapong, 2013, \Diamond from NE Makokou, Gabon. **8-9**. *Quamtana oku* Huber, 2003, \Diamond \Diamond with eggsacs from Mt. Oku, Cameroon. Photos BAH.

center of endemism until 2002, and numbers were similarly low for West Africa (19) and East Africa (29). This situation has changed dramatically within the last decade. Focused arachnological collecting campaigns by a number of individuals from various institutions have resulted in an increase of available



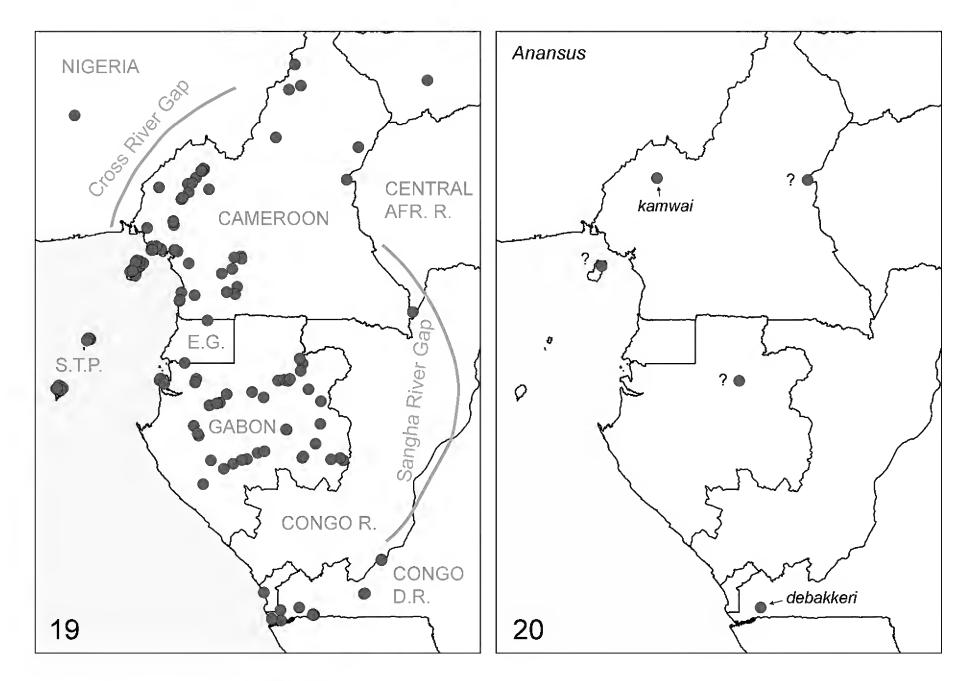
Figs 10-18. Litter, bark and ground-dwelling Central African pholcids. **10**. *Anansus kamwai* sp. nov., ♀ with eggsac from near Bamenda, Cameroon. **11**. *Pholcus moca* Huber, 2011, ♂ from near Ndjole, Gabon. **12-13**. *Pholcus punu* sp. nov., pair from between Mimongo and Yéno (12) and ♂ from Moudouma (13), Gabon. **14-15**. *Pholcus rawiriae* sp. nov., ♀ with eggsac (14) and ♂ from Mohoba, Gabon (15). **16**. *Smeringopina fang* Huber, 2013, ♀ with eggsac from Mayebout, Gabon. **17**. *Smeringopina belinga* Huber, 2013, ♂ from Monts de Belinga, Gabon. **18**. *Smeringopus lesserti* Kraus, 1957, ♂ from Mayebout, Gabon. Photos BAH.

material by about two to three orders of magnitude. Revisions of all major and most minor pholcid genera present in Africa have more than tripled the number of described African species (Huber 2003a, b, c, 2007, 2009, 2011b, 2012, 2013). Even though percentages of new species collected during intensive sampling trips suggest hundreds of undescribed species in tropical Africa (Huber in press), African pholcids can probably be considered fairly well known at the generic level. At least a framework is now available that roughly circumscribes each group in terms of morphology, distribution, and relationships, and that allows old species to be identified and new species to be recognized as such.

Two recent overviews of the Pholcidae from East Africa and West Africa have summarized the available information for each of these regions, including descriptions for species that had not been included in taxonomic revisions (Huber & Warui 2012; Huber & Kwapong 2013). The present paper is the last in this trilogy, concentrating of the third part of mainland tropical Africa that has received intensive recent attention both by collectors and in taxonomic revisions.

Material and methods

A large part of the material studied herein was collected during expeditions to Cameroon (2009) and Gabon (2011). This material is currently deposited at the Zoologisches Forschungsmuseum Alexander Koenig, Bonn (ZFMK). Further material was borrowed from the following institutions: Natural History Museum, London (BMNH); California Academy of Sciences, San Francisco (CAS); Musée royal de



Figs 19-20. All pholcid records known from Central Africa (19) and known distribution of the genus *Anansus* Huber, 2007 in Central Africa (20). Question marks denote uncertain identifications (females from Bioko and Gabon and one poorly preserved male from eastern Cameroon). E.G. = Equatorial Guinea; S.T.P. = São Tomé and Príncipe.

l'Afrique Centrale, Tervuren (MRAC); National Museum of Natural History, Washington D.C. (USNM); and Zoological Museum, University of Copenhagen, Copenhagen (ZMUC).

Format, style of descriptions, and terminology are as in recent revisions (e.g., Huber 2011b, 2012, 2013). Measurements are in mm unless otherwise noted. The precision for eye measurements is ca. 5 μ m. Epigyna were cleared in a warm NaOH solution and stained with chlorazol black. Locality coordinates are in round brackets when copied from labels and original publications or when received directly from collectors, in square brackets when originating from some other source (such as online gazetteers, Google Earth, MRAC database, etc.).

The following abbreviations are used:

ALE = anterior lateral eyes

ALS = anterior lateral spinnerets

AME = anterior median eyes

a.s.l. = above sea level L/d = length/diameter

PME = posterior median eyes

The geographic area considered here (between 10°N and 7°S; between 6°E and 18°E; Fig. 19) encompasses mainly the Lower Guinean subregion of the Guineo-Congolian center of endemism (White 2001). The western and eastern limits of this subregion are somewhat controversial. In the west, the main current gap (Dahomey Gap; ~0-3°E) is not always congruent with distribution limits of taxa, which in many cases are further east (Cross River Basin of eastern Nigeria, ~8°E) (Dowsett-Lemaire & Dowsett 2001; Lawson & Klemens 2001; Maley 2001). In the east, both the Gabon-Congo watershed and the Sangha-Ubangui Rivers have been proposed as limits (Dowsett-Lemaire & Dowsett 2001). For the present overview, however, the exact western and eastern limits are fairly irrelevant since the pholcid faunas of Nigeria and Congo Republic remain largely unknown. The southern and northern limits were chosen to include western Congo D.R. (with the excellent collections from Luki Forest Reserve in MRAC) and all known pholcid records from Cameroon (no record from beyond 10°N is known to us).

Results

Class Arachnida Cuvier, 1812 Order Araneae Clerck, 1757 Family Pholcidae C.L. Koch, 1851

Annotated list of Central African genera and species

Pholcidae described from Central Africa as defined herein, with countries, references, and new records.

Anansus Huber, 2007

Anansus is endemic and widely distributed in tropical Africa (Huber 2007; Huber & Warui 2012; Huber & Kwapong 2013), with two nominal species currently known from Central Africa (Fig. 20). Females in this genus are difficult to distinguish, and female specimens from Bioko (Pico Basilé) in CAS and from Gabon (Ntenkélé) in ZFMK (Ar 11886) might represent further undescribed species.

1. Anansus debakkeri Huber, 2007. Congo D.R. (Huber 2007).

New records

 2007 (D. De Bakker, J.P. Michiels); $7 \circlearrowleft \circlearrowleft 6 \hookrightarrow \hookrightarrow + \text{juvs}$ (7 vials) in MRAC (224312-315 part, 319 part, 234844-845), same locality, fogging in primary rainforest, 25 Sep.-4 Oct. 2007 (D. De Bakker, J.P. Michiels); $5 \circlearrowleft \circlearrowleft 6 \circlearrowleft 4 \hookrightarrow \hookrightarrow (6 \text{ vials})$ in MRAC (222190 part, 223021, 223039, 223120 part, 223457 part, 223679 part), same locality, beating in old secondary rainforest, 17-23 Sep. 2007 (D. De Bakker, J.P. Michiels); $4 \circlearrowleft \circlearrowleft (7 \text{ vials})$ in MRAC (223588 part, 223605 part, 223628 part, 223649, 223746, 223804, 223835), same locality, beating in primary rainforest, 26 Sep.-3 Oct. 2007 (D. De Bakker, J.P. Michiels); $1 \circlearrowleft (7 \text{ vials})$ in MRAC (222019), same locality, sweeping along trail in primary rainforest, 11 Nov. 2006 (D. De Bakker, J.P. Michiels); $1 \circlearrowleft (7 \text{ in MRAC})$ in MRAC (222753), same locality, pitfalls in primary rainforest, 13-23 Sep. 2007 (D. De Bakker, J.P. Michiels).

2. Anansus kamwai sp. nov. (see below). Cameroon.

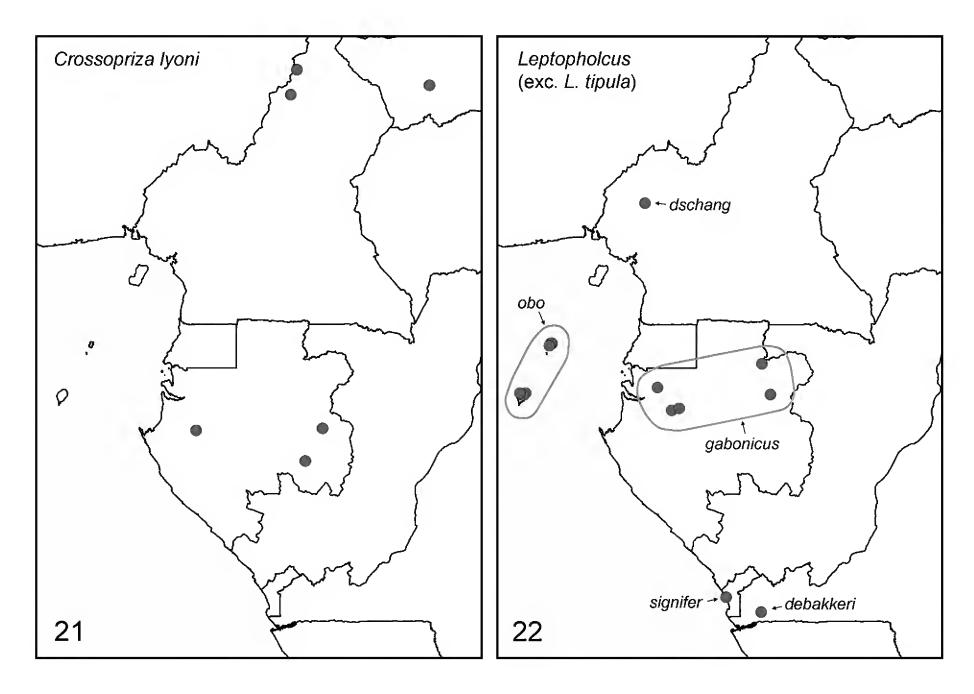
Crossopriza Simon, 1893

Only the pantropical synanthropic *C. lyoni* (Blackwall, 1867) occurs in Central Africa (Fig. 21). It is common but has apparently not been recorded previously from the area considered here.

3. Crossopriza lyoni (Blackwall, 1867). Newly recorded for Cameroon, Chad, and Gabon.

New records

CAMEROON: North Region: $2 \subsetneq \subsetneq$, 2 juvs in MRAC (148259), Touroua [9°04.7'N, 12°58.4'E], 8-9 Jan. 1976 (F. Puylaert). $4 \subsetneq \subsetneq$ in MRAC (221386), Faro Game Reserve [8°24'N, 12°49'E], in hut, 5 May 2007 (R. Jocqué *et al.*).



Figs 21-22. Known distributions of *Crossopriza lyoni* (Blackwall, 1867) and of the genus *Leptopholcus* Simon 1893 [except *L. tipula* (Simon, 1907); see Fig. 23] in Central Africa.

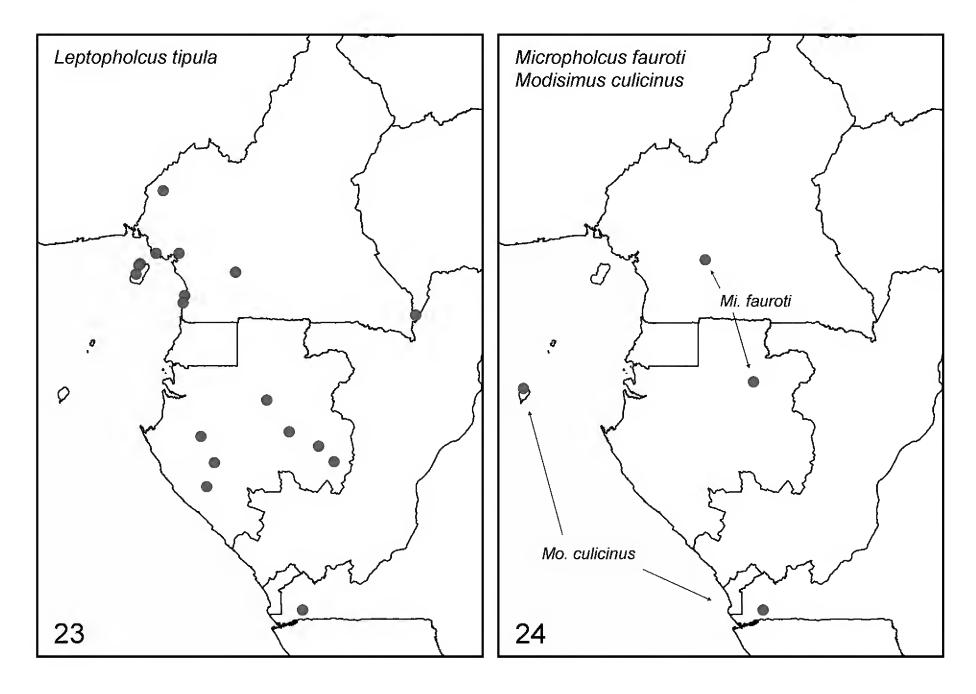
CHAD: Logone Oriental: $1 \circlearrowleft 3 \circlearrowleft 9 \circlearrowleft 3$ juvs in MRAC (151455), Bébédjia, near Moundou [8°40.6'N, 16°34.5'E], 1-10 Jul. 1977 (G. Ruella).

GABON: Haut Ogooué: 1 ♀ in ZFMK (Ar 11887), Mouanda (1°32.3'S, 13°12.6'E), 410 m a.s.l., in building, 20 Aug. 2011 (B.A. Huber). 2 ♂♂ in ZFMK (Ar 11888), Okondja (0°39'S, 13°40.5'E), 350 m a.s.l., in building, 17 Aug. 2011 (B.A. & S.R. Huber). Moyen-Ogooué: 1 ♂, 1 juv. in ZFMK (Ar 11889), Lambaréné (0°42.2'S, 10°14.4'E), 30 m a.s.l., at building, 27 Aug. 2011 (B.A. Huber).

Leptopholcus Simon, 1893

Of the ten species of *Leptopholcus* currently known from mainland Africa (Huber 2011b; Huber & Kwapong 2013), six occur in Central Africa (Figs 22-23). Of these, four are endemic to Central Africa (*L. dschang* Huber, 2011; *L. obo* Huber, 2011; *L. debakkeri* Huber, 2011; *L. gabonicus* sp. nov.); the fifth [*L. tipula* (Simon, 1907)] occurs throughout the entire Guineo-Congolian rainforest (Huber 2011b). The type species *L. signifer* Simon, 1893 was described from Central Africa (Cabinda, Angola) but its actual range is dubious (Huber 2011b).

- 4. Leptopholcus debakkeri Huber, 2011. Congo D.R. (Huber 2011b).
- 5. Leptopholcus dschang Huber, 2011. Cameroon (Huber 2011b).
- 6. Leptopholcus gabonicus sp. nov. (see below). Gabon.
- 7. Leptopholcus obo Huber, 2011. São Tomé and Príncipe (Huber 2011b).



Figs 23-24. Known distributions of *Leptopholcus tipula* (Simon, 1907), *Micropholcus fauroti* (Simon, 1887), and *Modisimus culicinus* (Simon, 1893) in Central Africa.

- 8. Leptopholcus signifer Simon, 1893. Angola (Cabinda) (Simon 1893).
- 9. *Leptopholcus tipula* (Simon, 1907). Equatorial Guinea (Bioko) (Simon 1907; Huber 2009); Cameroon, Central African Republic, Congo D.R., Gabon (Huber 2009).

New records

Micropholcus Deeleman-Reinhold & Prinsen, 1987

Only the synanthropic *M. fauroti* (Simon, 1887) occurs in Central Africa (Huber 2011b) (Fig. 24). It is probably more common than suggested by the few records.

10. Micropholcus fauroti (Simon, 1887). Cameroon (Huber 2011b); newly recorded for Gabon.

New record

GABON: Ogooué-Ivindo: $9 \circlearrowleft \circlearrowleft , 2 \hookrightarrow \circlearrowleft$ in ZFMK (Ar 11894), Makokou (0°33.9'N, 12°50.7'E), 520 m a.s.l., in building, 13 Aug. 2011 (B.A. & S.R. Huber).

Modisimus Simon, 1893

Of this otherwise New World genus, only the pantropical *M. culicinus* (Simon, 1893) occurs in Africa. It is probably common throughout the region, but rare in collections due to its small size and cryptic lifestyle.

11. *Modisimus culicinus* (Simon, 1893). Newly recorded for Central Africa [Fig. 24; Lessert's (1938) record - sub *Hedypsilus lawrencei* - from Kananga, Congo D.R., is outside the area considered here].

New records

SÃO TOMÉ AND PRÍNCIPE: São Tomé: 1 & in CAS, barricade wall S of Praia du Mutamba (0°23.2'N, 6°35.7'E), 3 May 2001 (J.M. Ledford).

CONGO D.R.: Bas Congo: 1 ♀ in MRAC (128843), Mayombe, Luki [5°39'S, 13°04'E], in *Cubitermes* [Termitidae] nest, 20 Sep. 1956 (A. Bouillon).

Ninetis Simon, 1890

This only African (and probably only Old World) genus of the subfamily Ninetinae is widely distributed from Namibia to the Arabian Peninsula, including Madagascar (Huber 2000, 2002; Huber & El Hennawy 2007). The new species described below is the first representative of the genus (and subfamily) recorded from Central Africa (Fig. 25).

12. Ninetis faro sp. nov. (see below). Cameroon.

Nyikoa Huber, 2007

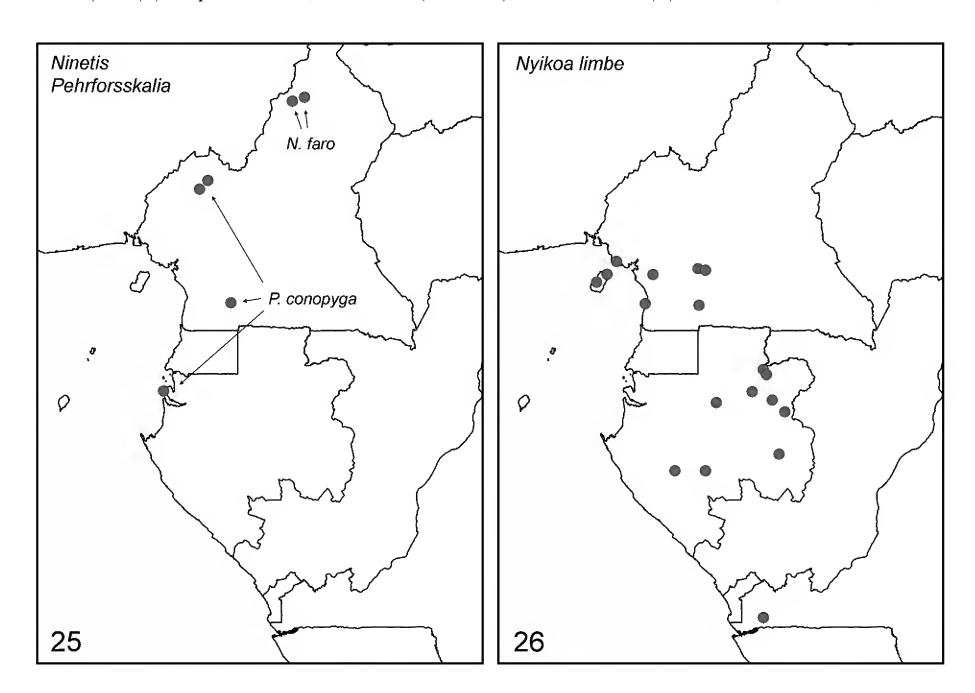
Nyikoa is a monotypic genus with the leaf-dwelling *N. limbe* Huber, 2007 widely distributed in the western subregions of the Guineo-Congolian rainforest (Upper and Lower Guinea) (Huber 2007, 2009; Huber & Kwapong 2013).

13. *Nyikoa limbe* Huber, 2007. Cameroon (Huber 2007, 2009), Congo D.R. (Huber 2007); newly recorded for Gabon and Equatorial Guinea (Fig. 26).

New records

CAMEROON: Central Region: 1 \(\pi \) in USNM, 6.5 km SSE of Yaounde (3°48.4'N, 11°32.4'E), Mbalmayo Rd., patch of forest, 715 m a.s.l., 8 Jan. 1992 (J. Coddington *et al.*).

EQUATORIAL GUINEA: Bioko: 2 \circlearrowleft in CAS, Pico Basilé (3°41.7'N, 8°52.3'E), ca. 700 m a.s.l., at night, 17 Oct. 1998 (D.K. Dabney, D. Ubick). 1 \circlearrowleft in CAS, 3.5 km N of Luba (3°28.9'N, 8°35.0'E), swampy forest, 13 Oct. 1998 (M. Boko *et al.*).



Figs 25-26. Known distributions of the genera *Ninetis* Simon, 1890 and *Pehrforsskalia* Deeleman-Reinhold & van Harten, 2001, and of *Nyikoa limbe* Huber, 2007 in Central Africa.

CONGO D.R.: Bas Congo: ~45 \circlearrowleft \circlearrowleft , 43 \circlearrowleft \circlearrowleft + juvs (8 vials) in MRAC (parts of 224303-307, 309-311), Luki Forest Reserve [5°37.3'S, 13°05.9'E], fogging in old secondary rainforest, 17-25 Sep. 2007 (D. De Bakker, J.P. Michiels); 76 \circlearrowleft \circlearrowleft , 47 \circlearrowleft \circlearrowleft + juvs (8 vials) in MRAC (parts of 224312-319), same data but fogging in primary rainforest, 26 Sept-4 Oct. 2007; 7 \circlearrowleft \circlearrowleft , 11 \circlearrowleft \circlearrowleft (6 vials) in MRAC (222207, 229, 994, 223096 part, 223123, 684), same locality, beating in old secondary rainforest, 17-25 Sep. 2007 (W. Hubau, D. De Bakker, J.P. Michiels); 12 \circlearrowleft \circlearrowleft , 13 \circlearrowleft \circlearrowleft , 2 juvs (9 vials) in MRAC (221533, 222026 part, 222061, 222798, 223635 part, 223587, 223732, 223819, 223831), same locality, beating in primary rainforest, 6-11 Nov. 2006, 26 Sep.-3 Oct. 2007 (D. De Bakker, J.P. Michiels); 1 \circlearrowleft in MRAC (221546), same locality, sieving, 7 Nov. 2006 (D. De Bakker, J.P. Michiels).

Pehrforsskalia Deeleman-Reinhold & van Harten, 2001

Pehrforsskalia includes three described species (Huber 2011b). Only the widely distributed P. conopyga Deeleman-Reinhold & van Harten, 2001 occurs in Central Africa (Fig. 25).

14. Pehrforsskalia conopyga Deeleman-Reinhold & van Harten, 2001. Cameroon, Gabon (Huber 2009).

Pholcus Walckenaer, 1805

Of the 29 operational species groups in *Pholcus* identified in Huber (2011b), only three occur in Central Africa: (1) the *bamboutos* group, with four of its nine species in Central Africa (Fig. 27; a fifth species, *P. kakum* Huber, 2009, is likely to occur in Lower Guinea but has not yet been documented from there); (2) the *circularis* group, which is largely endemic to the area considered here (Fig. 28; only *P. leruthi* occurs further west, in Congolia and East Africa); (3) the *debilis* group, which is largely endemic to the area considered here (Fig. 29; only *P. baka* ranges into Congolia). While the monophyly of the *debilis* group is well supported, the relationships within and among the other groups remain largely obscure (Huber 2011b; Dimitrov *et al.* 2013).

15. Pholcus attuleh Huber, 2011. Cameroon (Huber 2011b).

New record

CAMEROON: Southwest Region: 1 \(\pri\) in ZMUC, Mt. Koupé above Nyasoso (4°50'N, 9°41'E), forest, 1200-1600 m a.s.l., 16-19 Feb. 1992 (C.E. Griswold, N. Scharff, C. Wanzie, S. Larcher, Masongo).

16. Pholcus baka Huber, 2011. Cameroon, Central African Republic, Gabon (Huber 2011b).

New records

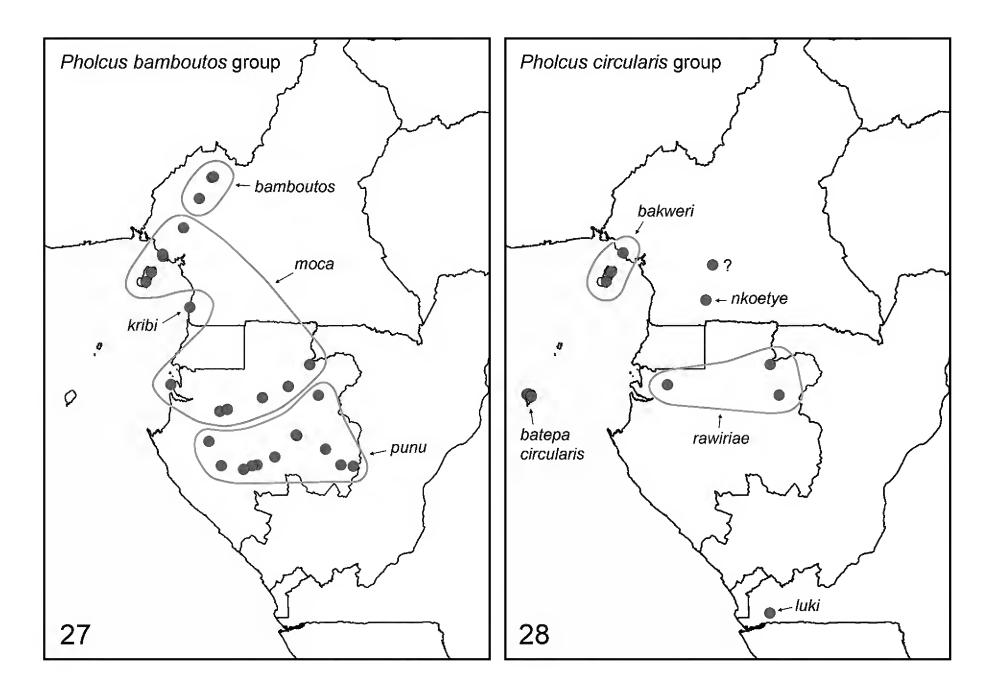
GABON: Ogooué-Ivindo: $1 \circlearrowleft 2 \circlearrowleft 1$ in ZFMK (Ar 11902), Monts de Belinga, forest near Mayebout (1°06.7'N, 13°06.6'E), 500 m a.s.l., 13-14 Aug. 2011 (B.A. & S.R. Huber); $1 \circlearrowleft 1$ in pure ethanol, in ZFMK (Gab 196), same data. $2 \circlearrowleft 2 \circlearrowleft 1$ in ZFMK (Ar 11903), Monts de Belinga, NE Makokou (0°47.0'N, 13°08.3'E), 530 m a.s.l., degraded forest along road, 15 Aug. 2011 (B.A. & S.R. Huber). $1 \circlearrowleft 1$ in ZFMK (Ar 11904), Ivindo N.P. near Simintang (0°32.2'N, 12°41.3'E), 545 m a.s.l., forest, 16

Aug. 2011 (B.A. & S.R. Huber). $1 \circlearrowleft$ in ZFMK (Ar 11905), near Djidji (0°12.8'N, 11°49.3'E), 470 m a.s.l., forest, 12 Aug. 2011 (B.A. & S.R. Huber). Haut Ogooué: $2 \circlearrowleft \circlearrowleft$, $1 \circlearrowleft$, 2 juvs in ZFMK (Ar 11906), forest above Canyon rose near Lékoni (1°38.6'S, 14°17.5'E), 600 m a.s.l., 19 Aug 2011 (B.A. & S.R. Huber); 1 juv. in pure ethanol, in ZFMK (Gab 210), same data. $9 \circlearrowleft \circlearrowleft$, $6 \circlearrowleft \circlearrowleft$ in ZFMK (Ar 11907), forest at Lékoni River (1°10.7'S, 13°32.3'E), 305 m a.s.l., 18 Aug. 2011 (B.A. & S.R. Huber); $1 \circlearrowleft$, 1 juv. in pure ethanol, in ZFMK (Gab 212), same data. $3 \circlearrowleft \circlearrowleft$, 1 juv. in ZFMK (Ar 11908), forest along Lékoni River (1°34.6'S, 14°12.1'E), 450 m a.s.l., 20 Aug. 2011 (B.A. & S.R. Huber). Moyen-Ogooué: $1 \circlearrowleft$, 1 \circlearrowleft in ZFMK (Ar 11909), S Lambaréné near Tchad (0°58.1'S, 10°22.7'E), 165 m a.s.l., forest, 27 Aug. 2011 (B.A. & S.R. Huber). $3 \circlearrowleft \circlearrowleft$, $3 \hookrightarrow \circlearrowleft$ + juvs in ZFMK (Ar 11910), near Ndjolé, 'site 1' (0°09.0'S, 10°40.0'E), 120 m a.s.l., forest near brook, 11 Aug. 2011 (B.A. & S.R. Huber); $1 \circlearrowleft$, $4 \hookrightarrow \circlearrowleft$, 1 juv. in pure ethanol, in ZFMK (Gab 228), same data. Estuaire: $2 \circlearrowleft \circlearrowleft$, $4 \hookrightarrow \circlearrowleft$, 2 juvs in ZFMK (Ar 11911), forest near Sahoué, N Libreville (0°34.6'N, 9°20.4'E), 30 m a.s.l., 28 Aug. 2011 (B.A. & S.R. Huber); $2 \hookrightarrow \circlearrowleft$ in pure ethanol, in ZFMK (Gab 181), same data.

- 17. Pholcus bakweri Huber, 2011. Cameroon, Equatorial Guinea (Huber 2011b).
- 18. Pholcus bamboutos Huber, 2011. Cameroon (Huber 2011b).

New record

CAMEROON: Northwest Region: 2 & d, 2 juvs in ZMUC, near Lake Oku (6°12'N, 10°27'E), forest at 2150 m a.s.l., 7-13 Feb. 1992 (C. Griswold, S. Larcher, N. Scharff, C. Wanzie).



Figs 27-28. Known distributions of the *bamboutos* and *circularis* species groups (*sensu* Huber 2011b) of *Pholcus* Walckenaer, 1805 in Central Africa. The question mark denotes an undescribed species represented by a single male in USNM.

- 19. Pholcus batepa Huber, 2011. São Tomé and Príncipe (Huber 2011b).
- 20. Pholcus circularis Kraus, 1957. São Tomé and Príncipe (Kraus 1957; Huber 2011b).
- 21. Pholcus debilis (Thorell, 1899). Cameroon, Equatorial Guinea (Bioko) (Thorell 1899; Huber 2011b).

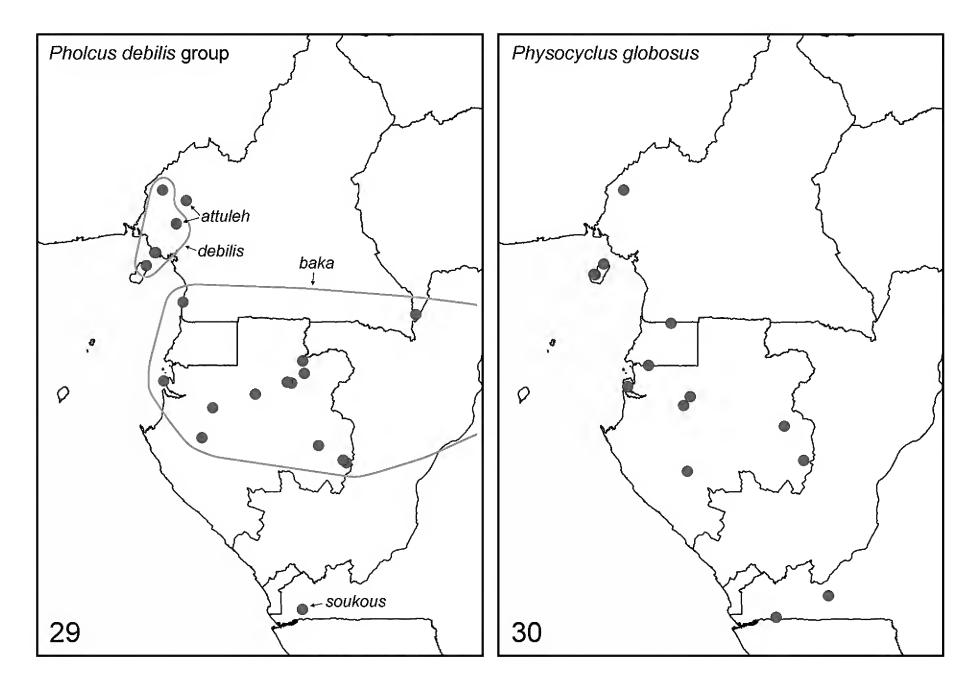
Note

The CAS has a single female specimen from Bioko (Pico Basilé) that may represent an additional (undescribed) species within the *debilis* group *sensu* Huber (2011b).

- 22. Pholcus kribi Huber, 2011. Cameroon (Huber 2011b).
- 23. Pholcus luki Huber, 2011. Congo D.R. (Huber 2011b).
- 24. *Pholcus moca* Huber, 2011. Cameroon, Equatorial Guinea (Bioko) (Huber 2011b); newly recorded from Gabon.

New records

GABON: Ogooué-Ivindo: $1 \, \circlearrowleft$, $2 \, \circlearrowleft \varphi$ in ZFMK (Ar 11912), Monts de Belinga, forest near Mayebout (1°06.7'N, 13°06.6'E), 500 m a.s.l., 13-14 Aug. 2011 (B.A. & S.R. Huber). $1 \, \circlearrowleft$ in ZFMK (Ar 11913), near Djidji (0°12.8'N, 11°49.3'E), 470 m a.s.l., forest, 12 Aug. 2011 (B.A. & S.R. Huber); $1 \, \circlearrowleft$, 1 juv. in pure ethanol, in ZFMK (Gab 240), same data. $1 \, \circlearrowleft$ in pure ethanol, in ZFMK (Gab 237), near Ntenkélé (0°31.4'N, 12°31.5'E), 550 m a.s.l., forest, 12. Aug. 2011 (B.A. & S.R. Huber). Moyen-Ogooué: $1 \, \circlearrowleft$, $2 \, \circlearrowleft$ in ZFMK (Ar 11914), near Ndjolé, 'site 1' (0°09.0'S, 10°40.0'E), 120 m a.s.l., forest near brook, 11 Aug. 2011 (B.A. & S.R. Huber). $2 \, \circlearrowleft$ in ZFMK (Ar 11915), near Ndjolé, 'site 2' (0°05.9'S,



Figs 29-30. Known distributions of the *debilis* species group (*sensu* Huber 2011b) of *Pholcus* Walckenaer, 1805 and of *Physocyclus globosus* (Taczanowski, 1874) in Central Africa.

10°52.4'E), 140 m a.s.l., forest near brook, 11 Aug. 2011 (B.A. & S.R. Huber). Estuaire: $1 \subsetneq$ in ZFMK (Ar 11916), forest near Sahoué, N Libreville (0°34.6'N, 9°20.4'E), 30 m a.s.l., 28 Aug. 2011 (B.A. & S.R. Huber); $1 \subsetneq$, 2 juvs in pure ethanol in ZFMK (Gab 180), same data.

25. Pholcus nkoetye Huber, 2011. Cameroon (Huber 2011b).

Note

The USNM has a single male specimen of a very similar undescribed species from Cameroon, SSE of Yaounde (Mbalmayo Road, 3°48.4'N, 11°32.4'E).

- 26. Pholcus punu sp. nov. (see below). Gabon.
- 27. Pholcus rawiriae sp. nov. (see below). Gabon.
- 28. Pholcus soukous Huber, 2011. Congo D.R. (Huber 2011b).

Physocyclus Simon, 1893

Physocyclus is a New World genus. Only the pantropical synanthropic *P. globosus* (Taczanowski, 1874) occurs in Central Africa (Fig. 30). It seems that this widespread species has not previously been recorded from the area considered here. Only Simon's (1877) *Pholcus claviger* (transferred and synonymzed in Huber 2011b) from "Congo" might be a previous record.

29. *Physocyclus globosus* (Taczanowski, 1874). Newly recorded for Cameroon, Equatorial Guinea, and Gabon.

New records

CAMEROON: Southwest Region: $1 \, \mathcal{J}$, $2 \, \mathcal{P}$, 3 juvs (2 vials) in BMNH, Mamfe [5°45'N, 9°19'E], in houses in clearing in forest, 13 Mar.-30 Apr. 1933 (Percy Sladen Trust Exped., I.T. Sanderson). Unidentified locality: $1 \, \mathcal{J}$, $1 \, \mathcal{P}$, 2 juvs (2 vials) in CAS, "Mabete Victoria Div.", 24 May-7 June 1949 (B. Malkin).

CONGO D.R.: Bas Congo: 4 \$\int 3\hat\cap , juvs (13 vials) in MRAC (1170, 1206-13, 26833, 26835-45, 26847, 26848, 26850, 26851, 26886-89, 26916, 26917, 26918, 26920), Matadi [5°49.3'S, 13°28.0'E], Feb. 1937 (E. Dartevelle). 1 \$\overline{\cap}\$ in MRAC (74564), Thysville [=Mbanza-Ngungu; 5°15'S, 14°52'E, ~600 m a.s.l.], 1953 (J. Sion).

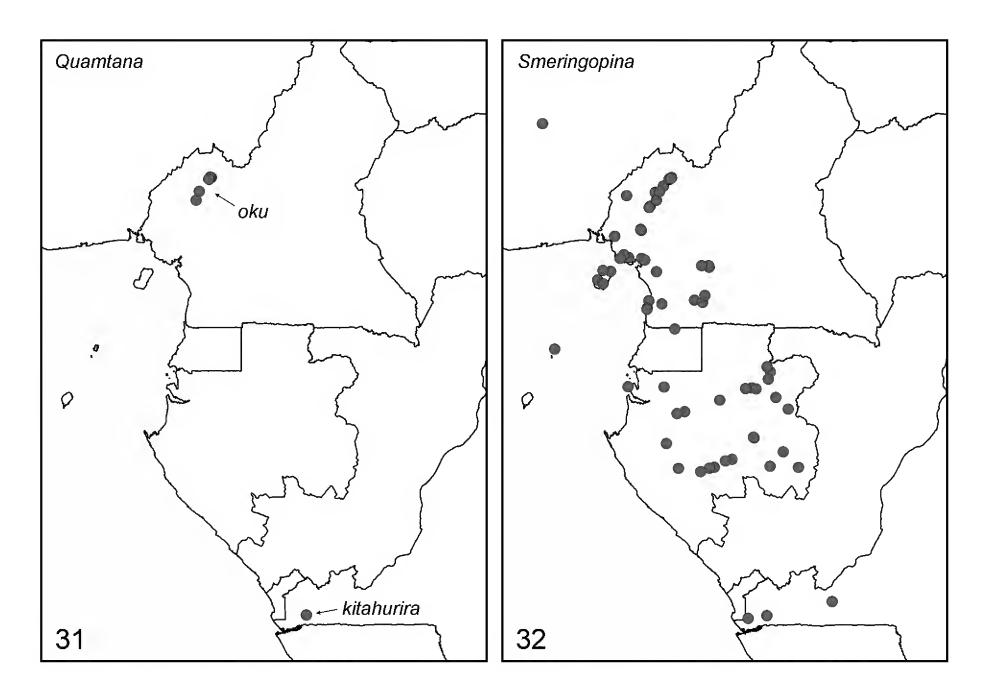
Quamtana Huber, 2003

Quamtana is highly diverse in southern Africa, but a few species occur further north in tropical Africa (Huber 2003c; Huber & Warui 2012; Huber & Kwapong 2013). In Central Africa, only Q. oku Huber, 2003 from Cameroon was previously known. The widespread Q. kitahurira Huber, 2003 (previously known from West and East Africa and from Angola) was expected to occur in Central Africa and is here newly reported for the region (Fig. 31).

30. Quamtana kitahurira Huber, 2003. Newly recorded for Central Africa.

New records

31. Quamtana oku Huber, 2003. Cameroon (Huber 2003c).



Figs 31-32. Known distributions of the genera *Quamtana* Huber, 2003 and *Smeringopina* Kraus, 1957 in Central Africa. For detailed distribution maps of *Smeringopina* species see Huber (2013).

New records

Smeringopina Kraus, 1957

The Guineo-Congolian genus *Smeringopina* has its highest diversity in Central Africa (Huber 2013). Any Central African forest seems to contain one to three representatives of the genus, and places without known records have most probably never been searched adequately (Fig. 32). Of the 44 currently known species, 37 occur in the area considered here. Except for *S. fon* Huber, 2013 (which ranges into West Africa) all of them are endemic to Central Africa.

- 32. Smeringopina africana (Thorell, 1899). Cameroon (Thorell 1899; Huber 2013).
- 33. Smeringopina armata (Thorell, 1899). Cameroon (Thorell 1899; Huber 2013).
- 34. Smeringopina attuleh Huber, 2013. Cameroon (Huber 2013).
- 35. Smeringopina bamenda Huber, 2013. Cameroon (Huber 2013).
- 36. Smeringopina bayaka Huber, 2013. Gabon (Huber 2013).
- 37. Smeringopina belinga Huber, 2013. Gabon (Huber 2013).
- 38. Smeringopina bioko Huber, 2013. Equatorial Guinea (Bioko) (Huber 2013).
- 39. Smeringopina bwiti Huber, 2013. Gabon (Huber 2013).
- 40. Smeringopina camerunensis Kraus, 1957. Cameroon (Kraus 1957; Huber 2013).
- 41. Smeringopina chaillu Huber, 2013. Gabon (Huber 2013).
- 42. Smeringopina cornigera (Simon, 1907). Cameroon (Simon 1907; Huber 2013).
- 43. Smeringopina djidji Huber, 2013. Gabon (Huber 2013).
- 44. Smeringopina ebolowa Huber, 2013. Cameroon (Huber 2013).
- 45. Smeringopina essotah Huber, 2013. Cameroon (Huber 2013).
- 46. Smeringopina etome Huber, 2013. Cameroon (Huber 2013).
- 47. Smeringopina fang Huber, 2013. Gabon (Huber 2013).
- 48. Smeringopina fon Huber, 2013. Nigeria, São Tomé and Príncipe (Huber 2013).
- 49. Smeringopina iboga Huber, 2013. Gabon (Huber 2013).

- 50. Smeringopina kala Huber, 2013. Cameroon, Equatorial Guinea (Huber 2013).
- 51. Smeringopina kikongo Huber, 2013. Congo D.R. (Huber 2013).
- 52. Smeringopina kinguele Huber, 2013. Gabon (Huber 2013).
- 53. Smeringopina kribi Huber, 2013. Cameroon (Huber 2013).
- 54. Smeringopina lekoni Huber, 2013. Gabon (Huber 2013).
- 55. Smeringopina luki Huber, 2013. Congo D.R. (Huber 2013).
- 56. Smeringopina mayebout Huber, 2013. Gabon (Huber 2013).
- 57. Smeringopina mbouda Huber, 2013. Cameroon (Huber 2013).
- 58. Smeringopina mohoba Huber, 2013. Gabon (Huber 2013).
- 59. Smeringopina moudouma Huber, 2013. Gabon (Huber 2013).
- 60. Smeringopina ndjole Huber, 2013. Gabon (Huber 2013).
- 61. Smeringopina ngungu Huber, 2013. Congo D.R. (Huber 2013).
- 62. Smeringopina nyasoso Huber, 2013. Cameroon (Huber 2013).
- 63. Smeringopina ogooue Huber, 2013. Gabon (Huber 2013).
- 64. Smeringopina sahoue Huber, 2013. Gabon (Huber 2013).
- 65. Smeringopina simintang Huber, 2013. Gabon (Huber 2013).
- 66. Smeringopina simplex Kraus, 1957. Cameroon (Kraus 1957; Huber 2013).
- 67. Smeringopina tchimbele Huber, 2013. Gabon (Huber 2013).
- 68. Smeringopina tebe Huber, 2013. Gabon (Huber 2013).

Smeringopus Simon, 1890

Smeringopus is currently the most species-rich African pholcid genus (55 species). It is widespread in Central Africa (Fig. 33), but only seven species are known from the area: the pantropical synanthropic *S. pallidus* (Blackwall, 1858), which is common in the area but whose previous records from Central Africa are all dubious; the widespread *S. cylindrogaster* (Simon, 1907), which covers at least the western subregions of the Guineo-Congolian rainforest (Upper and Lower Guinea) (Huber 2012), and its close relative *S. luki* Huber, 2012; the widespread *S. lesserti* Kraus, 1957, which covers at least the eastern subregions of the Guineo-Congolian rainforest (Lower Guinea and Congolia) (Huber 2012); and the three species of the *thomensis* group (*S. thomensis* Simon, 1907; *S. mayombe* Huber, 2012; *S. principe* Huber, 2012), all of which are small scale endemics (Huber 2012).

69. *Smeringopus cylindrogaster* (Simon, 1907). Cameroon (Huber 2009), Gabon (Huber 2012). Newly recorded for Equatorial Guinea (Bioko) and Central African Republic.

New records

EQUATORIAL GUINEA: Bioko: 1 \circlearrowleft in CAS, Arena Blanca, 7 km N of Luba (3°31.3'N, 8°35'E), 14 Oct. 1998 (M. Boko *et al.*).

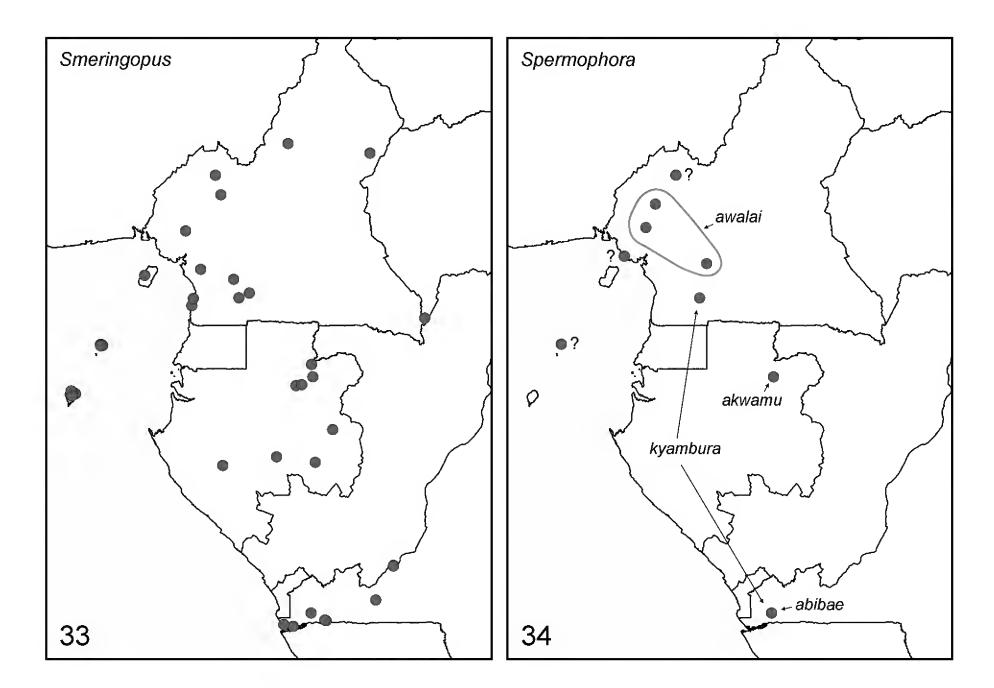
CENTRAL AFRICAN REPUBLIC: Sangha-Mbare: $3 \circlearrowleft 1$ juv. in CAS (9027099), Parc National Dzanga-Ndoki (2°22.2'N, 16°10.3'E), 360 m a.s.l., rainforest, beating low vegetation, 20-28 May 2001 (B.L. Fisher).

- 70. *Smeringopus lesserti* Kraus, 1957. Congo D.R. (Lessert 1930; Kraus 1957; Huber 2012), Cameroon, Gabon, São Tomé and Príncipe (Huber 2012).
- 71. Smeringopus luki Huber, 2012. Congo D.R. (Huber 2012).
- 72. Smeringopus mayombe Huber, 2012. Congo D.R. (Huber 2012).
- 73. Smeringopus pallidus (Blackwall, 1858). Congo D.R. (Simon 1877, sub *Pholcus excavatus*; see Huber 2011b); Congo R. (Lawrence 1958; doubtful record, only juveniles); newly recorded for Cameroon and Gabon.

New records

CAMEROON: Adamawa: ~5 \circlearrowleft \circlearrowleft , 20 \circlearrowleft \circlearrowleft , juvs (2 vials) in MRAC (141342, 343), Galim [7°05.8'N, 12°28.5'E], 13-20 Aug. 1971 (F. Puylaert). ~8 \circlearrowleft \circlearrowleft , 13 \circlearrowleft \circlearrowleft , juvs (2 vials) in MRAC (141390, 392), Djohong [6°50.2'N, 14°41.6'E], 1-5 Sep. 1971 (F. Puylaert). West Region: 1 \circlearrowleft in MRAC (148377), Kounden [5°42.2'N, 10°40.0'E], 28 Dec. 1975 (F. Puylaert).

GABON: Haut Ogooué: $1 \circlearrowleft$ in ZFMK (Ar 11927), Mouanda (1°32.3'S, 13°12.6'E), 410 m a.s.l., in building, 21 Aug. 2011 (B.A. Huber). $3 \circlearrowleft \circlearrowleft$, $5 \circlearrowleft \circlearrowleft$, 1 juv. in ZFMK (Ar 11928), Okondja (0°39'S, 13°40.5'E), 350 m a.s.l., in building, 17 Aug. 2011 (B.A. & S.R. Huber). Ogooué-Ivindo: $1 \circlearrowleft$, $2 \circlearrowleft \circlearrowleft$ in ZFMK (Ar 11929), Makokou (0°33.9'N, 12°50.7'E), 520 m a.s.l., in building, 15 Aug. 2011 (B.A. & S.R. Huber); $1 \circlearrowleft$, 1 juv. in pure ethanol, in ZFMK (Gab 218), same data.



Figs 33-34. Known distributions of the genera *Smeringopus* Simon, 1890 and *Spermophora* Hentz, 1841 in Central Africa. For detailed distribution maps of *Smeringopus* species see Huber (2012). Question marks denote uncertain identifications (females from Cameroon; one pair from Príncipe Island).

CONGO D.R.: Kinshasa: 1♂ in MRAC (25673), "Leopoldville" [=Kinshasa, 4°20'S, 15°19'E], Jul. 1945 (Bureau); 2♀♀, 1 juv. in MRAC (59225-30), same locality, Feb. 1947 (E. Dartevelle). 1♀ in MRAC (26834), Matadi [5°49.3'S, 13°28.0'E], Feb. 1937 (E. Dartevelle).

- 74. Smeringopus principe Huber, 2012. São Tomé and Príncipe (Príncipe) (Huber 2012).
- 75. Smeringopus thomensis Simon, 1907. São Tomé and Príncipe (São Tomé) (Simon 1907; Huber 2012).

Spermophora Hentz, 1841

With currently 25 species, *Spermophora* is quite diverse in sub-Saharan Africa and Madagascar, but only the widespread *S. kyambura* Huber & Warui, 2012 has previously been reported from Central Africa (Huber & Warui 2012). Three additional species are here reported from Central Africa (Fig. 34). Of these, *S. abibae* sp. nov. and *S. akwamu* Huber & Kwapong, 2013 are very similar to each other and to the West African *S. dieke* Huber, 2009. *Spermophora awalai* sp. nov. appears most similar to a group of East African species (node 5 in Appendix 4 in Huber 2003a). A further undescribed species seems to occur on São Tomé and Príncipe (NW of Santo Antonio, $1 \, \circlearrowleft$, $1 \, \circlearrowleft$ deposited in CAS). Females from Cameroon (Mt. Etinde, $1 \, \circlearrowleft$ in USNM; near Oku, $1 \, \circlearrowleft$ in pure ethanol in ZFMK, Cam 117) might represent further undescribed species.

- 76. Spermophora abibae sp. nov. (see below). Congo D.R.
- 77. Spermophora akwamu Huber & Kwapong, 2013. Newly recorded for Central Africa.

New record

GABON: Ogooué-Ivindo: $3 \circlearrowleft \circlearrowleft 6 \circlearrowleft 9 \circlearrowleft 3$ juvs in ZFMK (Ar 11930), Monts de Belinga, NE of Makokou (0°47.0'N, 13°08.3'E), 530 m a.s.l., degraded forest along road, 15 Aug. 2011 (B.A. & S.R. Huber); same data, 4 juvs in pure ethanol, in ZFMK (Gab 222).

Note

The new specimens are tentatively assigned to this species because the palps appear identical and the clypeus apophyses are at most minimally different (Figs 95-96). There is some slight difference in the color pattern but this has also been found among different populations in Ghana (Huber & Kwapong 2013).

- 78. Spermophora awalai sp. nov. (see below). Cameroon.
- 79. Spermophora kyambura Huber & Warui, 2012. Cameroon, Congo D.R. (Huber & Warui 2012).

Identification key to Central African pholcid genera

References to figures in upper case ("Fig.") refer to illustrations herein; figures in lower case ("fig.") have been published previously and are freely accessible online (at http://www.pholcidae.de/). Artema atlanta Walckenaer, 1837 is included as it probably occurs in Central Africa, even though we know of no records.

	Sternum with characteristic pattern of radiating lines (Fig. 36); abdomen dorsally with dark heart-mark (Fig. 35)
-	Sternum without or with different pattern; abdomen dorsal pattern different4
	Carapace with 2-3 pairs of dark lateral spots; ocular area (especially in males) strongly elevated, in male with frontal hairy pocket; male chelicerae without proximal lateral projections (figs 2-4 in Huber 1996)
	projections
	Male palpal bulb with three processes: embolus, uncus, appendix (Fig. 69). Epigynum sclerotized (at least posterior rim), with knob-shaped structure (Fig. 64) <i>Pholcus</i> Walckenaer, 1805 (part) Male palpal bulb with only one or two processes (embolus, bulbal apophysis); epigynum unsclerotized, without knob-shaped structure
	Procursus with highly complex system of transparent lamellae (fig. 14 in Huber 2007); male chelicerae narrowing distally, with pair of simple frontal apophyses close to median line (fig. 15 in Huber 2007) Nyikoa limbe Huber, 2007 Procursus and male chelicerae different Spermophora Hentz, 1841
	Carapace with median indentation (furrow or pit)
	Abdomen globular, oval, or higher than long
	Procursus with prominent distal spine; female carapace with posterior median cone acting against frontal plate on abdomen; epigynum with median anterior process
1(Legs with many small black marks; abdomen angular in lateral view (posteriorly high)
11	Male chelicerae with proximal lateral projections, either without or with several modified hairs on each side
12	Abdomen globular or oval
	Procursus very short and simple (Fig. 58), male chelicerae without proximal lateral apophyses (Fig. 60); epigynum sclerotized
	4 Procursus with long dorsal hinged process, female genitalia with internal U-shaped structure visible through cuticle anteriorly (figs 83-89 in Huber 2011b) <i>Micropholcus fauroti</i> (Simon, 1887)

- Procursus widely curved, without long dorsal process (figs 200, 205 in Huber 2003c); epigynum weakly sclerotized, no internal structure visible through cuticle Quamtana Huber, 2003
- Abdomen drawn into cone dorso-posteriorly (fig. 29 in Huber 2011b); male chelicerae with pair of lateral unsclerotized projections in distal position (fig. 130 in Huber 2009); epigynum weakly sclerotized Pehrforsskalia conopyga Deeleman-Reinhold & van Harten, 2001

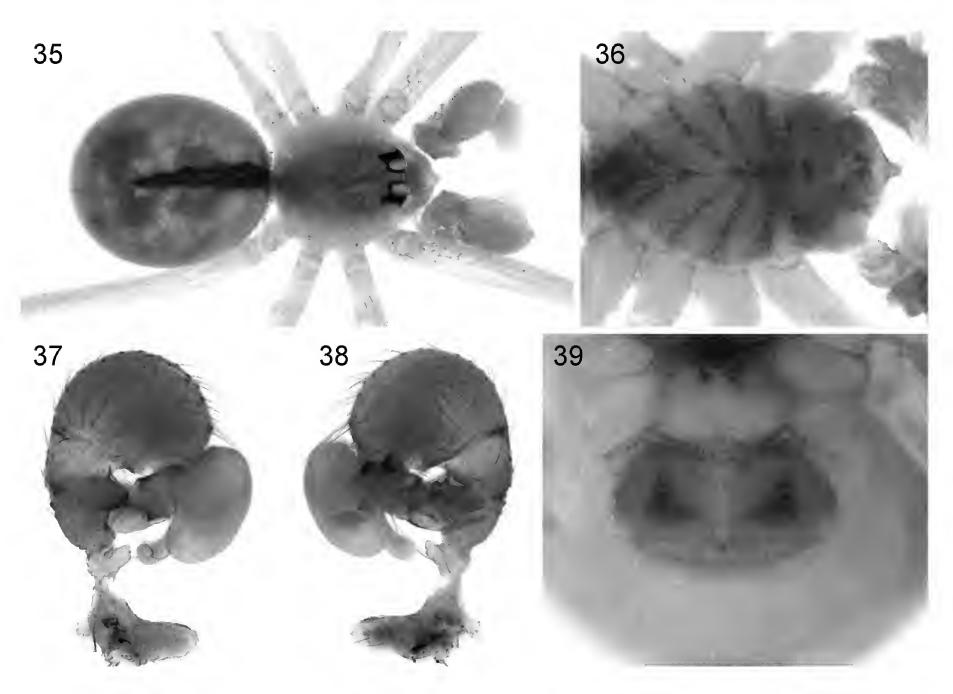
Taxonomy

Anansus kamwai Huber sp. nov.

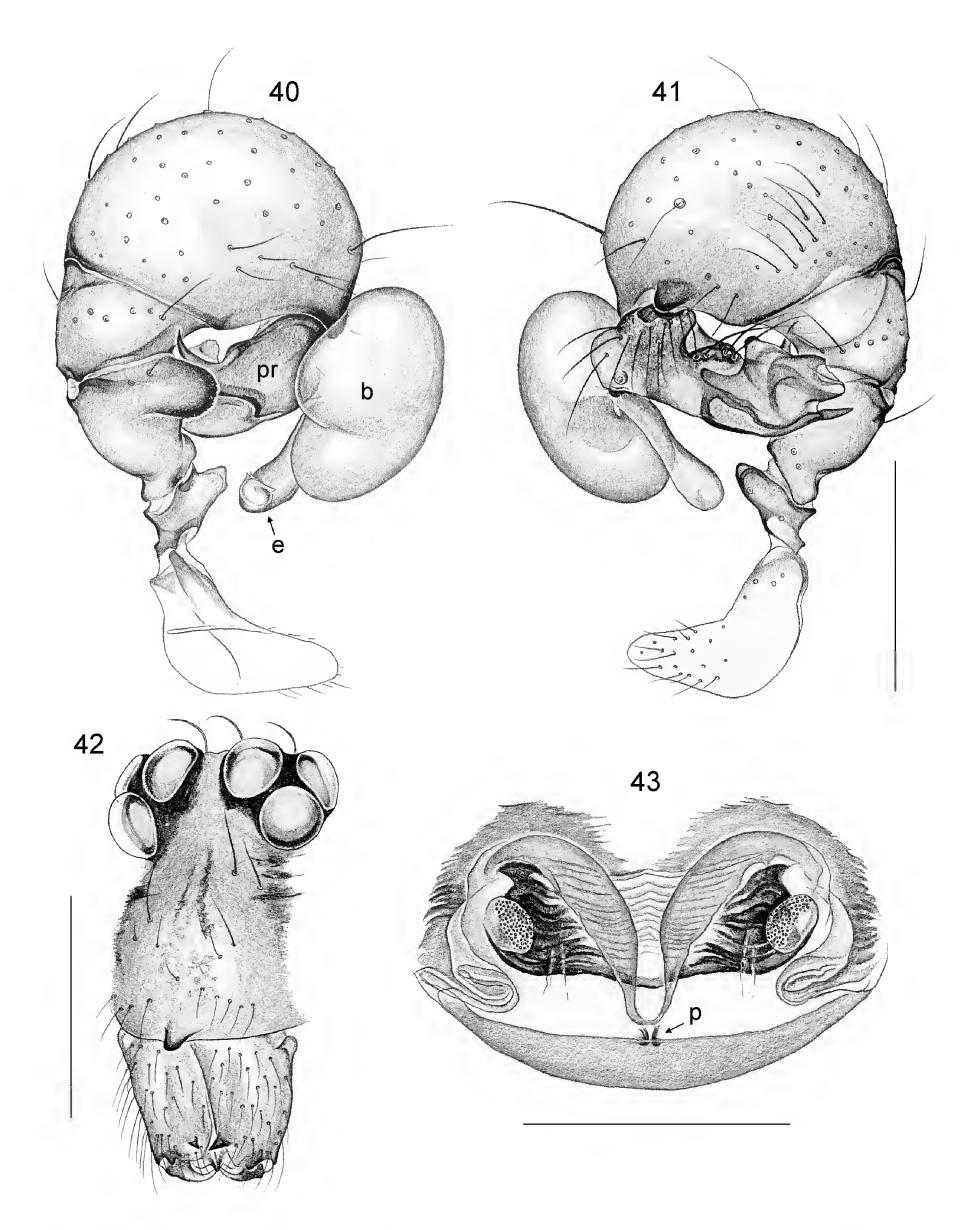
<u>urn:lsid:zoobank.org:act:716ECD2B-4969-4FF5-9A36-FA7CD47F9B08</u> Figs 10, 20, 35-43

Diagnosis

Distinguished from known congeners by shape of procursus (Figs 40, 41; distinctive distal elements); from *A. atewa* also by unpaired process on male clypeus (Figs 36, 42; paired in *A. atewa*); from other species also by more pointed apophyses frontally on male chelicerae (Fig. 42). Females in this genus are difficult to distinguish (except *A. atewa*, which has the epigynal pockets more anteriorly).



Figs 35-39. Anansus kamwai sp. nov. 35. \triangleleft , dorsal view. 36. \triangleleft sternum, ventral view. 37-38. Left \triangleleft palp, prolateral and retrolateral views. 39. \triangleleft epigynum, ventral view.



Figs 40-43. Anansus kamwai sp. nov. 40-41. Left \Im palp, prolateral and retrolateral views. 42. \Im ocular area, clypeus and chelicerae, oblique frontal view. 43. Cleared \Im genitalia, dorsal view. b = bulb; e = embolus; p = pocket; pr = procursus. Scale lines: 0.3 mm.

Etymology

Named for Cameroonian filmmaker Daniel Kamwa (born 1943), director of the 1981 film Notre Fille.

Type material

Holotype \circlearrowleft , in ZFMK (Ar 11931).

Type locality

CAMEROON, Northwest Region, near Bamenda, under trash and logs at roadside (6°00.5'N, 10°18.1'E), 1750 m a.s.l., 16 Apr. 2009 (B.A. Huber).

Other material examined

Description

Male (holotype)

Measurements. Total body length 1.6, carapace width 0.60. Leg 1: 6.00 (1.50 + 0.20 + 1.63 + 1.90 + 0.77), tibia 2: 1.07, tibia 3: 0.80, tibia 4: 1.20; tibia 1 L/d: 23. Distance PME-PME 45 μ m, diameter PME 60 μ m, distance PME-ALE 20 μ m; AME absent.

Color. Carapace ochre-yellow with irregular internal black pigment, sternum light brown with median dark line and four pairs of posteriorly diverging lines (Fig. 36); legs ochre-yellow without distinct dark rings; abdomen ochre-gray, with distinct dark heart-mark and thin dark line above spinnerets, dorsally with some white internal spots.

Body. Habitus as in Fig. 35; ocular area not elevated; carapace without median furrow; clypeus with rounded median process at rim (Fig. 42); sternum wider than long (0.48/0.36), unmodified. Chelicerae as in Fig. 42, with pair of lateral processes proximally and pair of pointed distal frontal apophyses without modified hairs; without stridulatory ridges.

Palps. As in Figs 37-38 and 40-41, coxa unmodified, trochanter with short ventral apophysis, femur small, with prominent distal apophysis prolaterally, tibia very large, procursus complex, with pointed ventral process, sclerotized retrolateral area set with several stronger hairs, distinctive tip; bulb with only one process (embolus, curved in dorsal view towards prolaterally).

Legs. Without spines and curved hairs, few vertical hairs; retrolateral trichobothrium on tibia 1 at 27%; prolateral trichobothrium absent on tibia 1, present on other tibiae; tarsus 1 with ~10 pseudosegments.

Variation. Tibia 1 in two other males: 1.37, 1.67.

Female

In general similar to male but clypeus unmodified; one female with darker chelicerae, clypeus, and sternum; tibia 1: 1.63 (missing in second female). Epigynum simple wide plate with pair of tiny pockets close together near posterior rim (Fig. 39), internal structures visible through cuticle; internal genitalia as in Fig. 43.

Distribution

Known from type locality in Cameroon only (Fig. 20). A poorly preserved male specimen from eastern Cameroon (Mbam near Koupoupi, MRAC 167.984) may also belong to this species.

Leptopholcus gabonicus Huber, sp. nov.

urn:lsid:zoobank.org:act:3CA8906E-7688-405E-AEDE-0A365C40281A Figs 1, 22, 44-52

Diagnosis

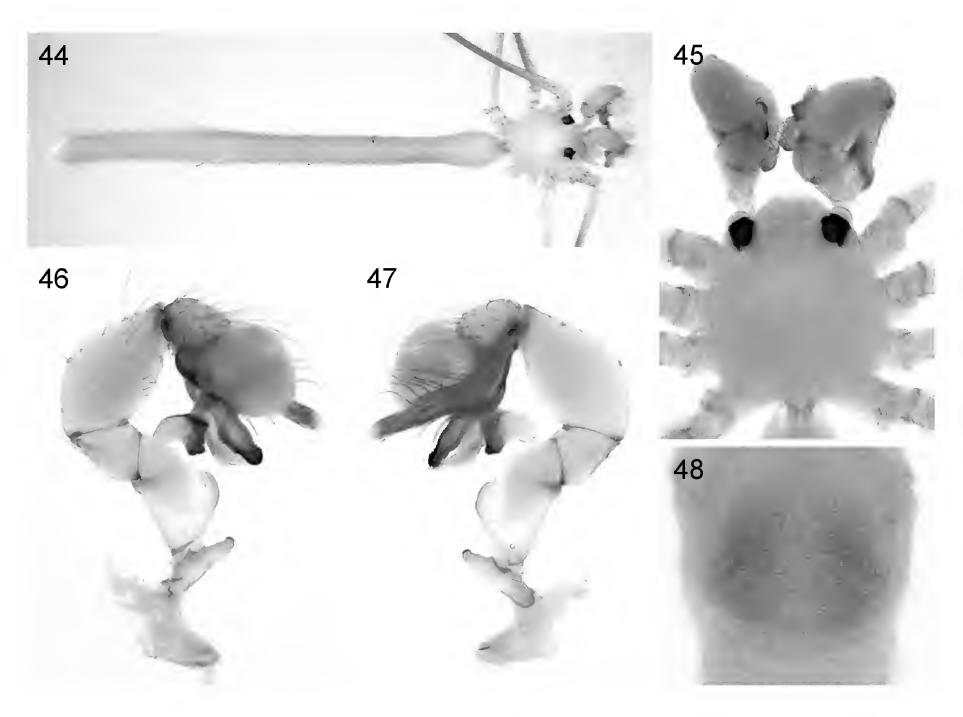
Distinguished from only other *Leptopholcus* species known from Gabon [*L. tipula* (Simon, 1907); cf. figs 82-86 in Huber 2009] by shapes of uncus (Fig. 49; much wider in *L. tipula*), with narrower semitransparent flap), appendix (Fig. 49; short and wide in *L. tipula*), and procursus (Fig. 50; wider tip in *L. tipula*); females of these two species are not easily distinguished. Distinguished from *L. obo* Huber, 2011 (São Tomé and Príncipe) by shapes of trochanter apophysis, procursus, uncus, appendix, and longer epigynum (cf. figs 324-327 in Huber 2011b); from other Central African species (*L. dschang* Huber, 2011; *L. signifer* Simon, 1893; *L. debakkeri* Huber, 2011) by absence of clypeus modification in male (Fig. 45).

Etymology

Named for the country of Gabon, where this species is endemic.

Type material

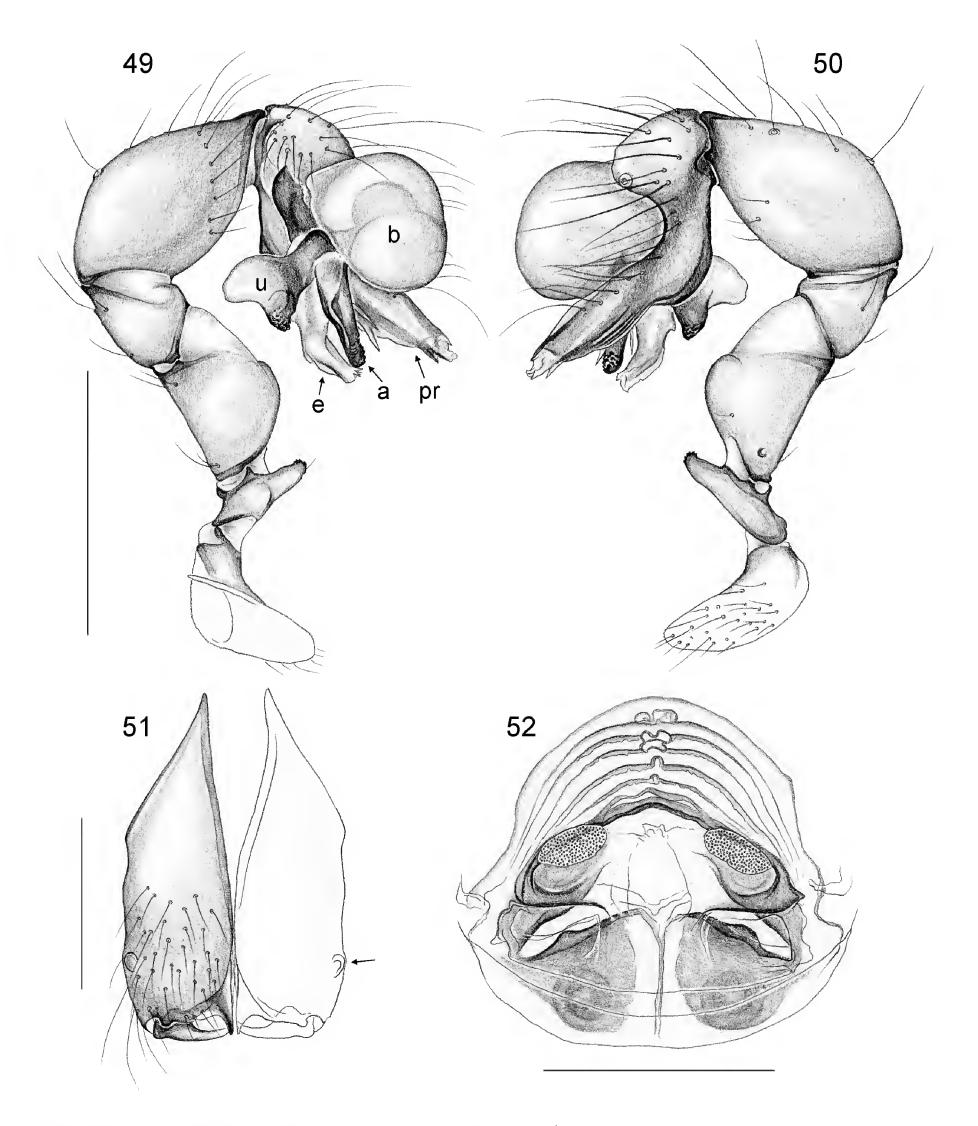
Holotype ♂, in ZFMK (Ar 11932).



Figs 44-48. Leptopholcus gabonicus sp. nov. 44. \circlearrowleft , dorsal view. 45. \circlearrowleft prosoma and palps, dorsal view. 46-47. Left \circlearrowleft palp, prolateral and retrolateral views. 48. \circlearrowleft epigynum, ventral view.

Type locality

GABON, Ogooué-Ivindo, Monts de Belinga, forest near Mayebout (1°06.7'N, 13°06.6'E), 500 m a.s.l., 13-14 Aug. 2011 (B.A. & S.R. Huber).



Figs 49-52. Leptopholcus gabonicus sp. nov. **49-50**. Left 3 palp, prolateral and retrolateral views. **51**. 3 chelicerae, frontal view (arrow points at lateral apophysis). **52**. Cleared 4 genitalia, dorsal view. 4 appendix; 4 b = 4 bulb; 4 e = 4 embolus; 4 procursus; 4 uncus. Scale lines: 4 constant 4 mm; 4 chelicerae 4 genitalia, dorsal view. 4 appendix; 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis). 4 chelicerae, frontal view (arrow points at lateral apophysis).

Other material examined

GABON: Ogooué-Ivindo: $3 \circlearrowleft \circlearrowleft 4 \circlearrowleft \varphi \hookrightarrow in ZFMK$ (Ar 11933), same data as holotype; $2 \circlearrowleft \varphi \hookrightarrow 3$ juvs in pure ethanol, in ZFMK (Gab 160, 200), same data. $1 \circlearrowleft in ZFMK$ (Ar 11934), near Mohoba Mozeye (0°16.8'N, 13°20.9'E), 510 m a.s.l., forest along road, 17 Aug. 2011 (B.A. & S.R. Huber); 1 juv. in pure ethanol, in ZFMK (Gab 170), same data. Moyen-Ogooué: $1 \circlearrowleft in ZFMK$ (Ar 11935), near Ndjolé, 'site 2' (0°05.9'S, 10°52.4'E), 140 m a.s.l., forest near brook, 11 Aug. 2011 (B.A. & S.R. Huber). $1 \hookrightarrow in ZFMK$ (Ar 11936), near Ndjolé, 'site 1' (0°09.0'S, 10°40.0'E), 120 m a.s.l., forest near brook, 11 Aug. 2011 (B.A. & S.R. Huber); 1 juv. in pure ethanol, in ZFMK (Gab 229), same data. Estuaire: $2 \circlearrowleft \circlearrowleft \circlearrowleft 1$ in ZFMK (Ar 11937), Monts de Cristal, near Kinguélé (0°27.8'N, 10°16.7'E), 100 m a.s.l., forest, 10 Aug. 2011 (B.A. & S.R. Huber); $2 \hookrightarrow \circlearrowleft 1$ in pure ethanol, in ZFMK (Gab 190), same data.

Description

Male (holotype)

Measurements. Total body length 6.0, carapace width 0.77. Leg 1: 33.4 (8.5 + 0.4 + 7.7 + 13.3 + 3.5), tibia 2: 5.8, tibia 3: 3.8, tibia 4: 6.4; tibia 1 L/d: 116. Distance PME-PME 290 μ m, diameter PME 80 μ m, distance PME-ALE 25 μ m, AME absent.

Color. Entire animal pale whitish to ochre-yellow, only patella area and tibia-metatarsus joints darker.

Body. Habitus as in Fig. 44; ocular area not elevated, each triad on low hump; carapace without median furrow; clypeus unmodified; sternum wider than long (0.64/0.46), unmodified. Chelicerae as in Fig. 51, with pair of small lateral processes barely visible in dissecting microscope; without stridulatory ridges.

Palps. As in Figs 46-47 and 49-50, coxa unmodified, trochanter with ventral apophysis with serrated tip, femur with large ventral hump and small retrolateral process proximally, procursus rather simple, with two membranous ventral processes, one arising from ventral 'knee', the other more distally; bulb with weakly sclerotized embolus, rod-shaped appendix with small scales, uncus consisting of sclerotized part with small scales and lighter dorsal flap.

Legs. Without spines and curved hairs, few vertical hairs; retrolateral trichobothrium on tibia 1 at 3%; prolateral trichobothrium absent on tibia 1, present on other tibiae; tarsus 1 pseudosegments not seen.

Variation. Tibia 1 in 6 other males: 7.1-7.7 (mean 7.3).

Female

In general similar to male; tibia 1 in 6 females: 6.2-7.0 (mean 6.6). Epigynum very simple, weakly sclerotized plate, without 'knob', internal structures visible through cuticle (Fig. 48); internal genitalia as in Fig. 52.

Natural history

All specimens were found on the undersides of leaves.

Distribution

Known from various localities in Gabon (Fig. 22).

Ninetis faro Huber, sp. nov.

urn:lsid:zoobank.org:act:C0D5E7FB-BB45-4921-BCBE-75FC4197FF02

Figs 25, 53-62

Diagnosis

Easily distinguished from most known congeners (especially from *N. subtilissima* Simon, 1890 which has a similar male palp) by shape of male cheliceral apophyses (Figs 60-61); from *N. russellsmithi* Huber, 2002 (which has similar male chelicerae) by shorter embolus and presence of procursus (Figs 58-59). From other species by shapes of bulbal projections (both projections of same length in *N. toliara* Huber & El Hennawy, 2007) or by shape of procursus (larger and not pointed in *N. minuta* (Berland, 1919) and *N. namibiae* Huber, 2000). Females are not easily distinguished (epigynum wider in *N. namibiae*, narrower in *N. minuta*; similar in other species).

Etymology

The name is a noun in apposition, derived from the type locality.

Type material

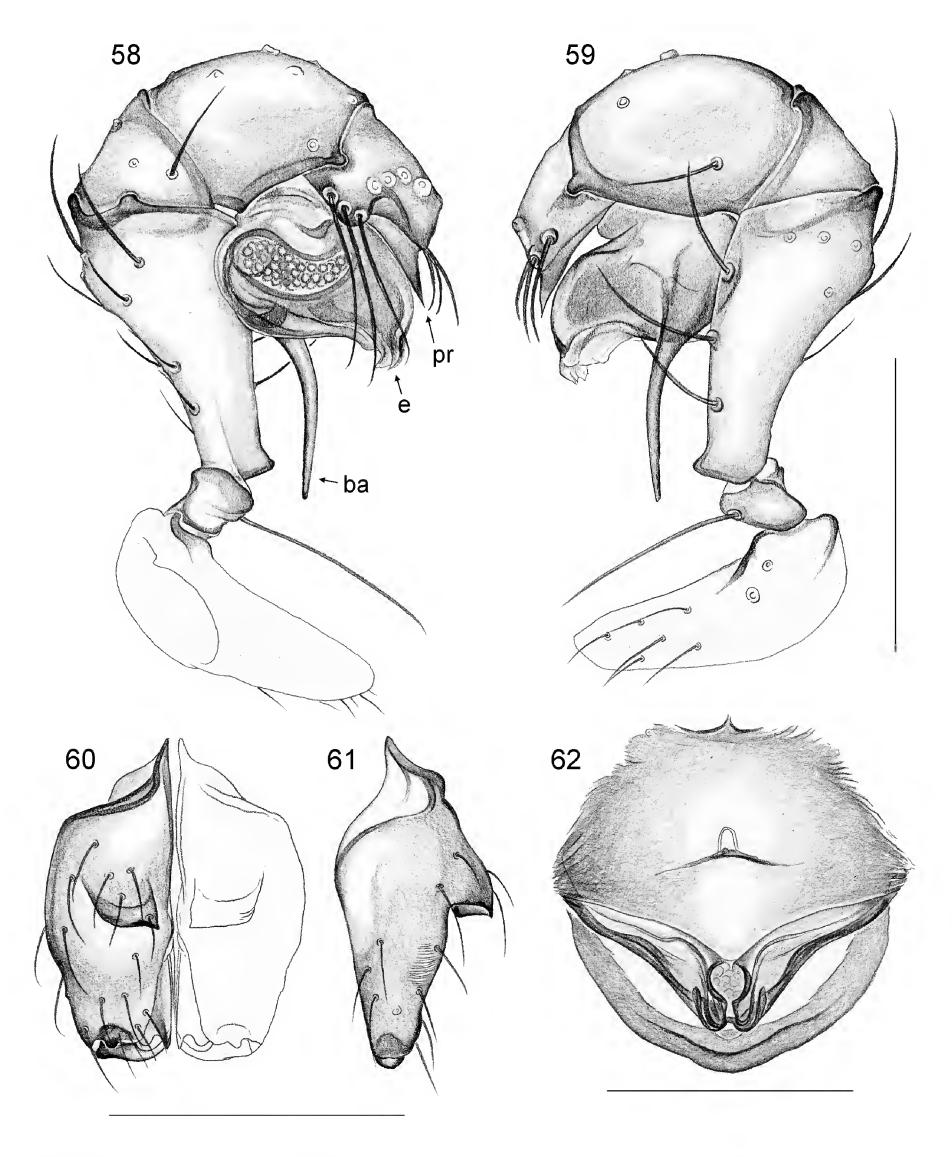
Holotype ♂, in MRAC (221178).

Type locality

CAMEROON, North Region, Faro Game Reserve [8°24'N, 12°49'E], pitfall in gallery forest, 5 May 2007 (R. Jocqué, K. Loosveldt, L. Baert, M. Alderweireldt).



Figs 53-57. *Ninetis faro* sp. nov. **53-54**. *△*, dorsal and lateral views. **55-56**. Left *△* palp, prolateral and retrolateral views. **57**. ♀, ventral view.



Figs 58-62. *Ninetis faro* sp. nov. **58-59**. Left \Diamond palp, prolateral and retrolateral views. **60-61**. \Diamond chelicerae, frontal and lateral views. **62**. Cleared \Diamond genitalia, dorsal view. ba = bulbal apophysis; e = embolus; pr = procursus. Scale lines: 0.2 mm.

Other material examined

CAMEROON: North Region: 1 \circlearrowleft in MRAC (221178), same data as and together with holotype; 2 \circlearrowleft \circlearrowleft 1 \circlearrowleft (2 vials) in MRAC (221136, 221208), same data but pitfall in wooded savanna, 3-4 May 2007. 1 \circlearrowleft in MRAC (221401), same data but sieving in gallery forest, 17 Apr. 2007. 2 \circlearrowleft \circlearrowleft 4 \circlearrowleft in MRAC (221463), Hossere Gare [8°30'N, 13°08'E], sieving of litter among rocks, 4 May 2007 (Jocqué, Loosveldt, Baert, Alderweireldt).

Description

Male (holotype)

Measurements. Total body length 1.2, carapace width 0.50. Leg 1: 2.96 (0.80 + 0.20 + 0.80 + 0.73 + 0.43), tibia 2: 0.67, tibia 3: 0.50, tibia 4: 0.80; tibia 1 L/d: 12. Distance PME-PME 35 µm, diameter PME 40 µm, distance PME-ALE 25 µm, diameter AME 20 µm, distance AME-AME 20 µm.

Color. Entire animal ochre-yellow to ochre-grey.

Body. Habitus as in Figs 53-54; ocular area not elevated; carapace without median furrow; clypeus more sclerotized at rim and medially slightly protruding; sternum about as wide as long (0.38/0.37), with distinct humps near leg coxae 1. Chelicerae as in Figs 60-61, with distinctive pair of frontal apophyses; with very fine stridulatory ridges.

PALPS. As in Figs 55-56 and 58-59, coxa and trochanter unmodified, femur with small retrolatero-ventral apophysis, tarsus with small pointed procursus; bulb with long ventral process and short membranous dorsal embolus.

Legs. Without spines and curved hairs, few vertical hairs; retrolateral trichobothrium on tibia 1 at 67%; prolateral trichobothrium apparently absent on tibia 1, present on other tibiae; tarsus 1 with ~6 pseudosegments.

Variation. Tibia 1 in 5 other males: 0.63-0.73 (mean 0.69).

Female

In general similar to male but clypeus, sternum, and chelicerae unmodified; tibia 1 in 5 females: 0.50-0.60 (mean 0.53). Epigynum simple oval sclerotized area consisting of large anterior and smaller posterior plate; internal structures poorly visible through cuticle (Fig. 57); unclear if complex internal structures are part of anterior plate (as appears in dorsal view; Fig. 62) or posterior plate (as appears in ventral view); anterior plate with transversal ridge apparently connected to median pocket. Pore plates not seen (Fig. 62).

Distribution

Known from two localities in the North Region of Cameroon (Fig. 25).

Pholcus punu Huber, sp. nov. <u>urn:lsid:zoobank.org:act:C721A4A8-5D18-467B-A433-7E526A641082</u> Figs 12-13, 27, 63-74

Diagnosis

Distinguished from *P. moca* Huber, 2011 (the most similar species and probably closest known relative) by shape of procursus (longer retrolateral process; absence of distinct ridge between retrolateral process and tip; compare Fig. 70 with fig. 946 in Huber 2011b); also by shape of uncus (not bilobed as in

P. moca), presence of proximal frontal processes on male chelicerae and slightly different shape of epigynal plate (narrower in *P. moca*; compare Fig. 64 with fig. 921 in Huber 2011b).

Etymology

Named for the Punu (or Bapunu, Bapounou), one of the four major peoples of Gabon; noun in apposition.

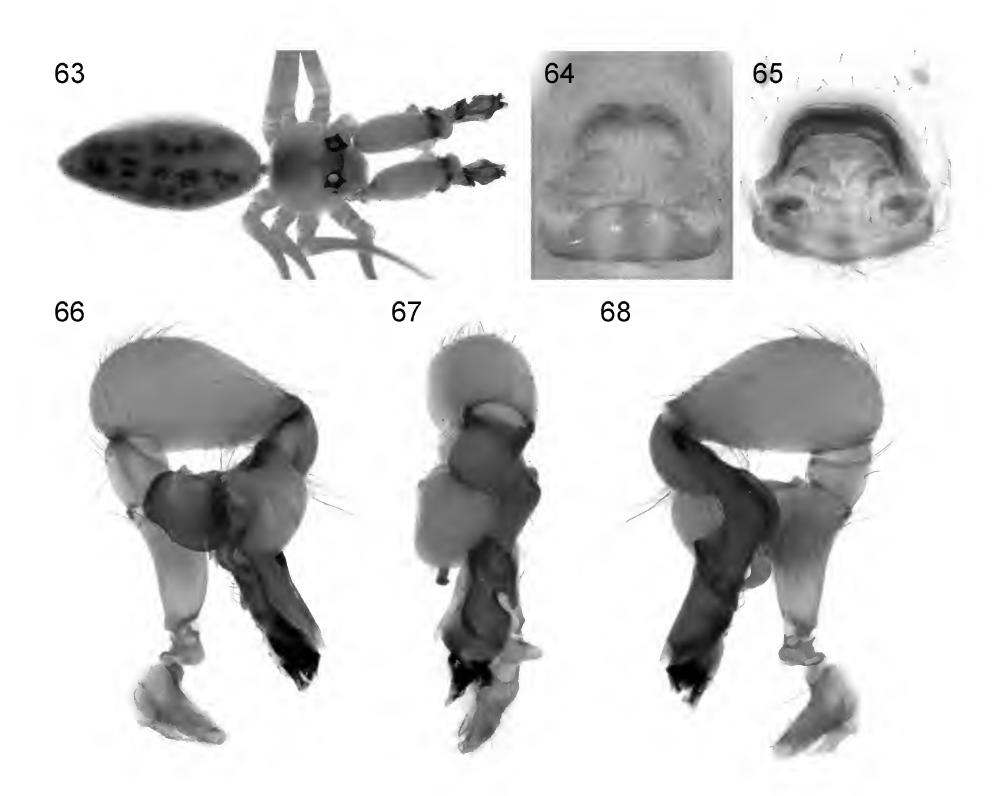
Type material

Holotype ♂, in ZFMK (Ar 11938).

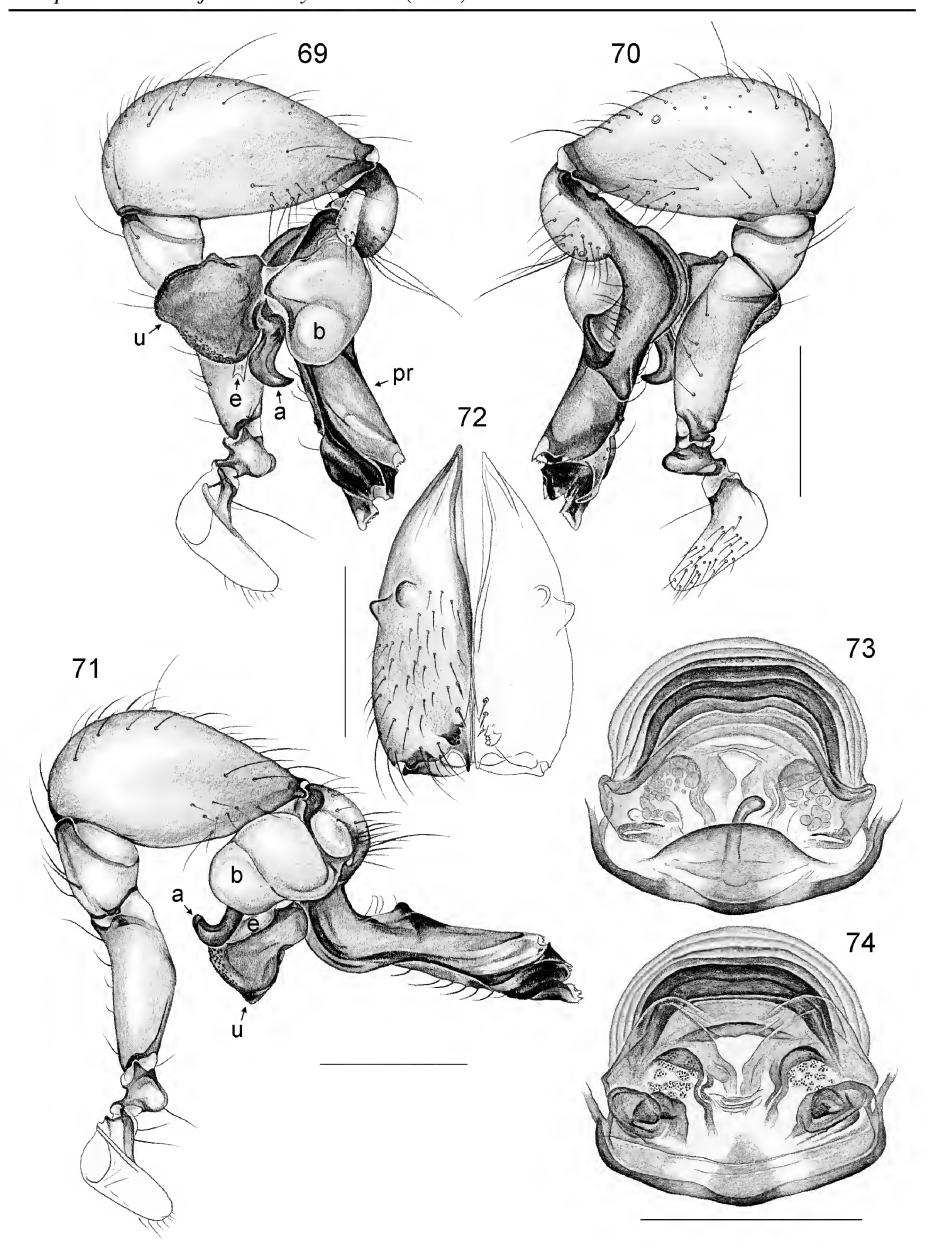
Type locality

GABON, Haut Ogooué, forest above Canyon rose near Lékoni (1°38.6'S, 14°17.5'E), 600 m a.s.l., 19 Aug. 2011 (B.A. & S.R. Huber).

Other material examined



Figs 63-68. *Pholcus punu* sp. nov. **63**. ♂, dorsal view. **64**. Epigynum, ventral view. **65**. Cleared ♀ genitalia, dorsal view. **66-68**. Left ♂ palp, prolateral, dorsal, and retrolateral views.



Figs 69-74. *Pholcus punu* sp. nov. **69-70**. Left \bigcirc palp, prolateral and retrolateral views. **71**. Extended palp with rotated bulb. **72**. \bigcirc chelicerae, frontal view. **73-74**. Cleared \bigcirc genitalia, ventral and dorsal views. a = appendix; b = bulb; e = embolus; pr = procursus; u = uncus. Scale lines: 69-71, 73-74 = 0.5 mm; 12 = 0.3 mm.

(Gab 213), same data. $1 \stackrel{>}{\circlearrowleft}$, $3 \stackrel{\frown}{\hookrightarrow}$ in ZFMK (Ar 11941), forest near Bongoville (1°36.7'S, 13°57.4'E), 600-650 m a.s.l., 20 Aug. 2011 (B.A. Huber); 2 juvs in pure ethanol, in ZFMK (Gab 207), same data. Ogooué-Ivindo: 5 ♂♂, 4 ♀♀ in ZFMK (Ar 11942), near Mohoba Mozeye (0°16.8'N, 13°20.9'E), 510 m a.s.l., forest along road, 17 Aug. 2011 (B.A. & S.R. Huber); 2 \(\Qmathbb{Q}\), 3 juvs in pure ethanol, in ZFMK (Gab 169), same data. Ogooué-Lolo: 1 ♂, 1 ♀ in ZFMK (Ar 11943), forest at brook near Lastoursville, near Grotte de Pahon Pira (0°48.8'S, 12°45.2'E), 290 m a.s.l., 22-23 Aug. 2011 (B.A. Huber). 2 ♂♂, 1 ♀ in ZFMK (Ar 11944), forest near Lastoursville (0°48.0'S, 12°44.4'E), 300 m a.s.l., 21 Aug. 2011 (B.A. Huber); 3 juvs in pure ethanol, in ZFMK (Gab 221), same data. 1 ♂, 1 ♀ in ZFMK (Ar 11945), near Moudouma (1°23.5'S, 12°09.6'E), 475 m a.s.l., forest along brook, 24 Aug. 2011 (B.A. & S.R. Huber); Massif du Chaillu, 'site 2', near Moukabou (1°36.6'S, 11°40.7'E), 560 m a.s.l., forest, 25 Aug. 2011 (B.A. & S.R. Huber); 3 juvs in pure ethanol, in ZFMK (Gab 174), same data. 4 \circlearrowleft 7, 7 \circlearrowleft 9 in ZFMK (Ar 11947), Massif du Chaillu, 'site 3', between Mimongo & Yéno (1°38.1'S, 11°32.6'E), 570-650 m a.s.l., forest, 26 Aug. 2011 (B.A. & S.R. Huber); 3 juvs in pure ethanol, in ZFMK (Gab 233), same data. 1 d in ZFMK (Ar 11948), Massif du Chaillu, 'site 4', between Yéno and Mouila (1°43.7'S, 11°18.4'E), 650 m a.s.l., forest along river, 26 Aug. 2011 (B.A. & S.R. Huber). 3 ♂♂, 7 ♀♀ in ZFMK (Ar 11949), near Moulandoufouala (1°38.1'S, 10°42.5'E), 110 m a.s.l., forest along road, 27 Aug. 2011 (B.A. & S.R. Huber); 4 juvs in pure ethanol, in ZFMK (Gab 184), same data. Moyen-Ogooué: 3 ♂♂ in ZFMK (Ar 11950), S of Lambaréné near Tchad (0°58.1'S, 10°22.7'E), 165 m a.s.l., forest, 27 Aug. 2011 (B.A. & S.R. Huber).

Description

Male (holotype)

Measurements. Total body length 3.0, carapace width 0.9. Leg 1: 27.0 (6.4 + 0.4 + 6.5 + 11.5 + 2.2), tibia 2: 4.5, tibia 3: 2.7, tibia 4: 4.1; tibia 1 L/d: 82. Distance PME-PME 265 μ m, diameter PME 95 μ m, distance PME-ALE 45 μ m, AME absent (only black spots without lenses).

COLOR. Carapace ochre-orange with large brown median mark, ocular area also brown, sternum medially slightly darkened; legs with dark rings on femora subdistally, patellae plus tibiae proximally, and tibiae subdistally; abdomen grey with internal black and whitish spots dorsally and laterally.

Body. Habitus as in Fig. 63; ocular area slightly elevated, each triad on short hump directed laterally; carapace without median furrow; clypeus unmodified; sternum wider than long (0.66/0.52), unmodified. Chelicerae as in Fig. 72, with pair of lateral processes proximally, pair of frontal apophyses proximally, and pair of frontal apophyses distally provided with two modified hairs each; without stridulatory ridges.

Palps. As in Figs 66-71, coxa unmodified, trochanter with short and wide retrolatero-ventral apophysis, femur with short retrolatero-dorsal process proximally, procursus complex, with distinctive sclerotized and membranous elements; bulb with large uncus, simple weakly sclerotized embolus, and hooked appendix.

LEGS. Without spines and curved hairs, few vertical hairs; retrolateral trichobothrium on tibia 1 at 3%; prolateral trichobothrium absent on tibia 1, present on other tibiae; tarsus 1 with >20 pseudosegments, only distally fairly distinct.

Variation. Tibia 1 in 22 other males: 6.7-8.8 (mean 7.9). Clypeus variably dark; AME spots sometimes fused. In most males the palps are extended and the bulbs rotated as in Fig. 71.

Female

In general similar to male; tibia 1 in 36 females: 5.3-7.1 (mean 6.3). Epigynum with distinct plate and 'knob', anterior area weakly sclerotized, 'valve' and other internal structures visible through cuticle (Fig. 64); internal genitalia as in Figs 65 and 73-74.

Natural history

The spiders were mostly found in the leaf litter and in cavities in the ground (Fig. 12), sometimes sharing the microhabitat with a representative of *Smeringopina* (e.g. *S. moudouma* Huber, 2013 in forest near Lastoursville; *S. fang* Huber, 2013 between Yéno and Mouila; *S. lekoni* Huber, 2013 at Lékoni River). The two specimens from Moudouma were found at the rock surface (Fig. 13) close to a small brook.

Distribution

Known from various localities in Gabon south of the Ogooué-Ivindo Rivers (Fig. 27; the closely related *P. moca* Huber, 2011 seems to be confined to the north of these rivers).

Pholcus rawiriae Huber, sp. nov.

urn:lsid:zoobank.org:act:54111E6C-353C-45DC-B72C-AC7D01382709 Figs 14-15, 28, 75-83

Diagnosis

Easily distinguished from known congeners by morphology of male palp (large trochanter apophysis; ventral femur apophyses; procursus with distal hinged sclerite strongly bent towards prolateral; bent embolus with spine-like processes; shape of appendix; Figs 80-81); from putative close relatives (bamboutos and circularis species groups) also by distinctive pattern on carapace (Fig. 75; transversal band on carapace; compare with figs. 892-918 in Huber 2011b), and by female external and internal genitalia (narrow sclerotized rim and distinctive shape of 'valve'; long pore plates; Figs 76, 83).

Etymology

Named for Gabonese novelist Angèle Ntyugwetondo Rawiri (1954–2010).

Type material

Holotype ♂, in ZFMK (Ar 11951).

Type locality

GABON, Ogooué-Ivindo, near Mohoba Mozeye (0°16.8'N, 13°20.9'E), 510 m a.s.l., forest along road, 17 Aug. 2011 (B.A. & S.R. Huber).

Other material examined

GABON: Ogooué-Ivindo: $2 \subsetneq \varphi$ in ZFMK (Ar 11951), same data as and together with holotype. $2 \circlearrowleft \Im$, $3 \hookrightarrow \varphi$ in ZFMK (Ar 11952), Monts de Belinga, forest near Mayebout (1°06.7'N, 13°06.6'E), 500 m a.s.l., 13-14 Aug. 2011 (B.A. & S.R. Huber); $2 \hookrightarrow \varphi$ in pure ethanol, in ZFMK (Gab 199), same data. Estuaire: $1 \hookrightarrow \Im$, 3 juvs in pure ethanol, in ZFMK (Gab 191), Monts de Cristal, between Tchimbélé and Kinguélé (0°33.8'N, 10°18.8'E), 520 m a.s.l., forest near river, 10 Aug. 2011 (B.A. & S.R. Huber).

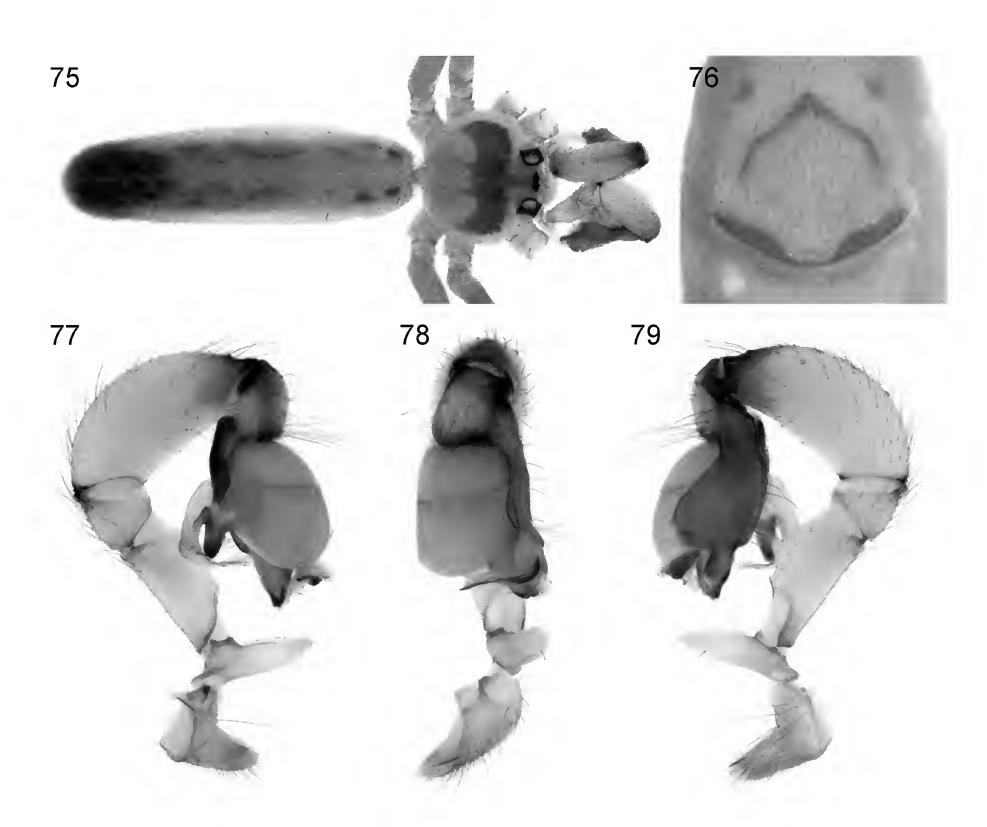
Description

Male (holotype)

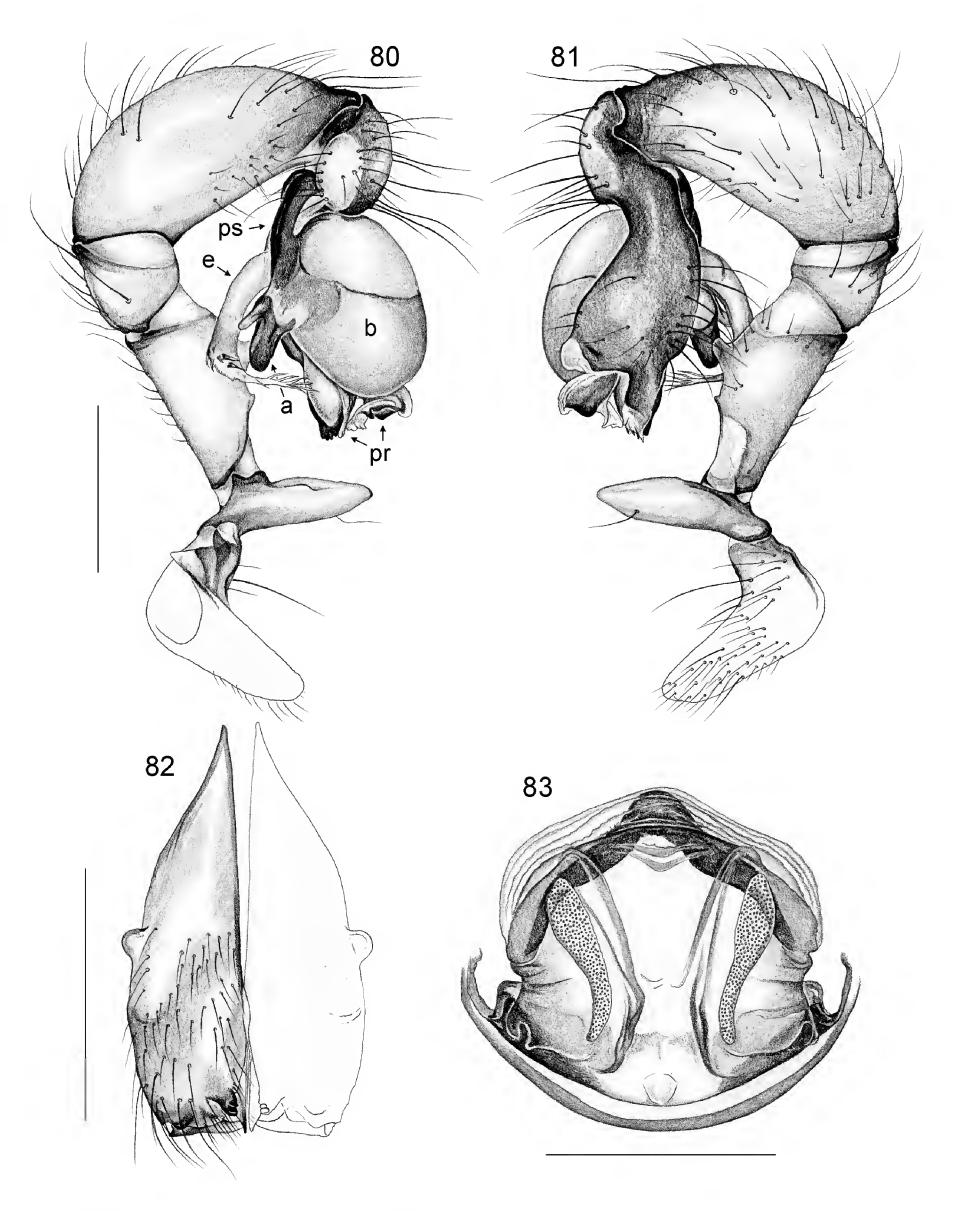
Measurements. Total body length 5.0, carapace width 1.4. Leg 1: 39.1~(9.6 + 0.5 + 9.3 + 17.5 + 2.2), tibia 2: 6.4, tibia 3: 4.2, tibia 4: 6.0; tibia 1 L/d: 62. Distance PME-PME 365 μ m, diameter PME 140 μ m, distance PME-ALE 45 μ m, diameter AME 90 μ m, distance AME-AME 70 μ m.

Color. Carapace ochre-yellow with distinctive dark pattern, clypeus not darkened, sternum with darker brown margins; legs with distinct dark rings on femora subdistally, patellae plus tibiae proximally, tibiae subdistally, and metatarsi proximally, tips of femora and tibiae lighter; abdomen dorsally and laterally with internal black marks.

Body. Habitus as in Fig. 75; ocular area slightly elevated, each triad on short hump directed laterally; carapace without median furrow; clypeus unmodified; sternum wider than long (0.92/0.70), unmodified. Chelicerae as in Fig. 82, with pair of small lateral processes proximally, directed slightly towards posterior, pair of low frontal humps, and pair of distal frontal apophyses provided with two modified hairs each; without stridulatory ridges.



Figs 75-79. *Pholcus rawiriae* sp. nov. **75**. ♂, dorsal view. **76**. ♀ epigynum, ventral view. **77-79**. Left ♂ palp, prolateral, dorsal, and retrolateral views.



Figs 80-83. Pholcus rawiriae sp. nov. 80-81. Left β palp, prolateral and retrolateral views. 82. β chelicerae, frontal view. 83. Cleared φ genitalia, dorsal view. α = appendix; α = bulb; α = embolus; α = procursus; α = proximal sclerite of bulb. Scale lines: 0.5 mm.

Palps. As in Figs 77-81, coxa unmodified, trochanter with very large ventral apophysis, weakly sclerotized and distally curved towards prolateral, femur with one small and one larger distinctive ventral projection, procursus with distinctive hinged process distally strongly curved towards prolateral; bulb with long proximal sclerite, strongly bent embolus provided with several spine-like processes and distal fringes, without uncus, appendix with weakly sclerotized basal process.

Legs. Without spines and curved hairs, few vertical hairs; retrolateral trichobothrium on tibia 1 at 4%; prolateral trichobothrium absent on tibia 1, present on other tibiae; tarsus 1 with >20 pseudosegments, only distally fairly distinct.

Variation. Not seen. Tibiae 1 missing in other males.

Female

In general similar to male; tibia 1 in 5 females: 6.6-8.1 (mean 7.2). Epigynum with narrow sclerotized rim posteriorly, median 'knob', anterior area weakly sclerotized, 'valve' visible through cuticle (Fig. 76); internal genitalia as in Fig. 83.

Natural history

Most specimens were found at tree trunks, with their bodies and legs tightly pressed against the bark and thus barely visible (Figs 14-15).

Distribution

Known from three localities in northern Gabon (Fig. 28).

Spermophora abibae Huber, sp. nov.

<u>urn:lsid:zoobank.org:act:85A657B7-245E-4943-881F-A86BEC8B934A</u> Figs 34, 84-96

Diagnosis

Distinguished from the very similar *S. dieke* Huber, 2009 and *S. akwamu* Huber & Kwapong, 2013 by bifid process on male clypeus (Figs 85, 92; only one median process in *S. dieke*; two processes in *S. akwamu*; cf. Figs 94-96), and by pore plates in female internal genitalia (Fig. 93; longer and in different position than in *S. dieke* and *S. akwamu*).

Etymology

The species name commemorates the 11-year old girl, who was purchased for six handkerchiefs by James Jameson, a member of Henry Stanley's Emin Pasha Relief Expedition, who gave her to cannibals so he could watch her being killed, cooked and eaten. Her name is not known. Abiba is an African name, meaning beloved.

Type material

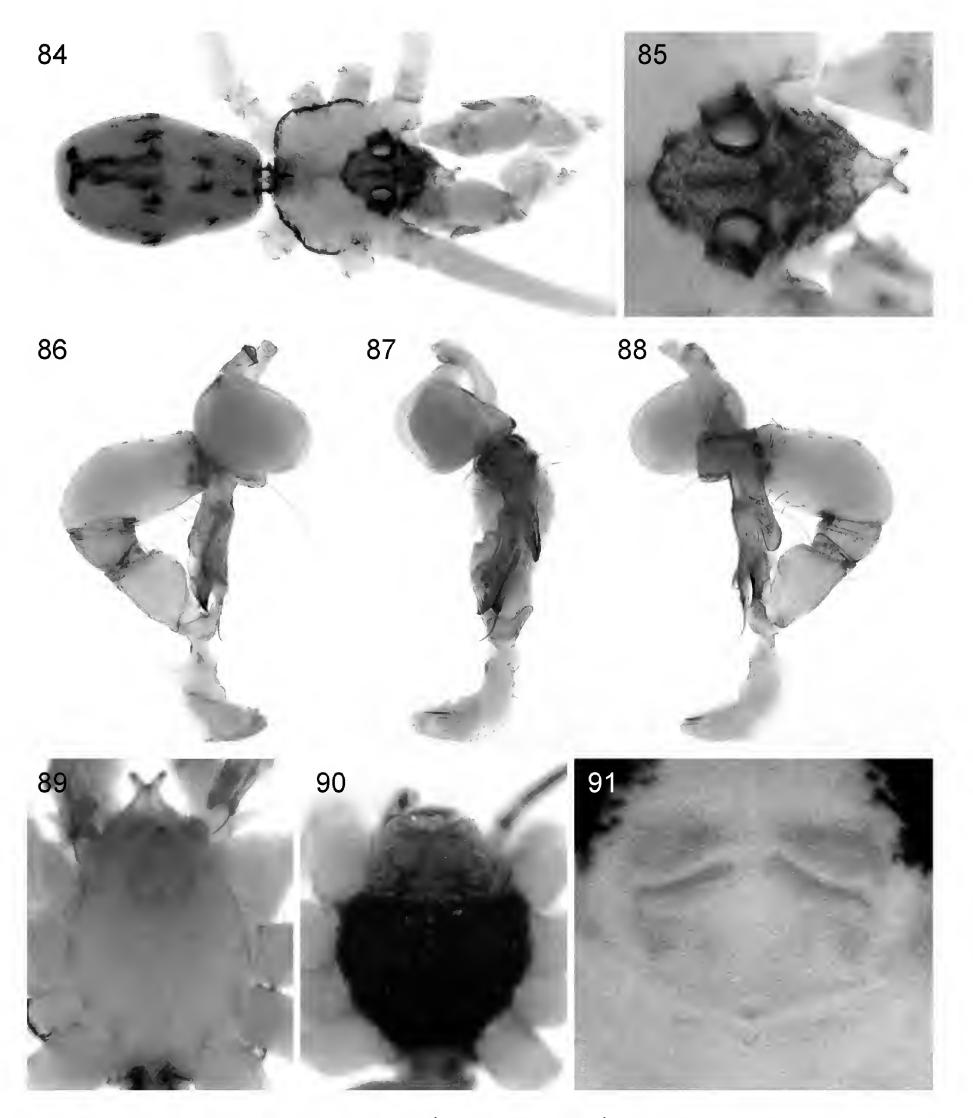
Holotype ♂, in MRAC (separated from 224303).

Type locality

CONGO D.R., Bas Congo, Mayombe, Luki Forest Reserve [5°37.3'S, 13°05.9'E], fogging in old secondary rainforest, 17 Sep. 2007 (D. De Bakker, J.P. Michiels).

Other material examined

CONGO D.R.: Bas Congo: $7 \circlearrowleft \circlearrowleft$, $9 \circlearrowleft \circlearrowleft$ in MRAC (224303 part), same data as holotype; $80 \circlearrowleft \circlearrowleft$, $107 \circlearrowleft \circlearrowleft$, juvs (8 vials) in MRAC (224304-11 part), same data but 18-25 Sep. 2007 (D. De Bakker, J.P. Michiels). $18 \circlearrowleft \circlearrowleft$, $29 \circlearrowleft \circlearrowleft$, juvs (11 vials) in MRAC (219850 part, 219852-54 part, 224312-17 part,



Figs 84-91. *Spermophora abibae* sp. nov. **84**. \triangleleft , dorsal view. **85**. \triangleleft ocular area and clypeus, dorsal view. **86-88**. Left \triangleleft palp, prolateral, dorsal, and retrolateral views. **89-90**. \triangleleft and \triangleleft prosomata, ventral views. **91**. \triangleleft epigynum, ventral view.

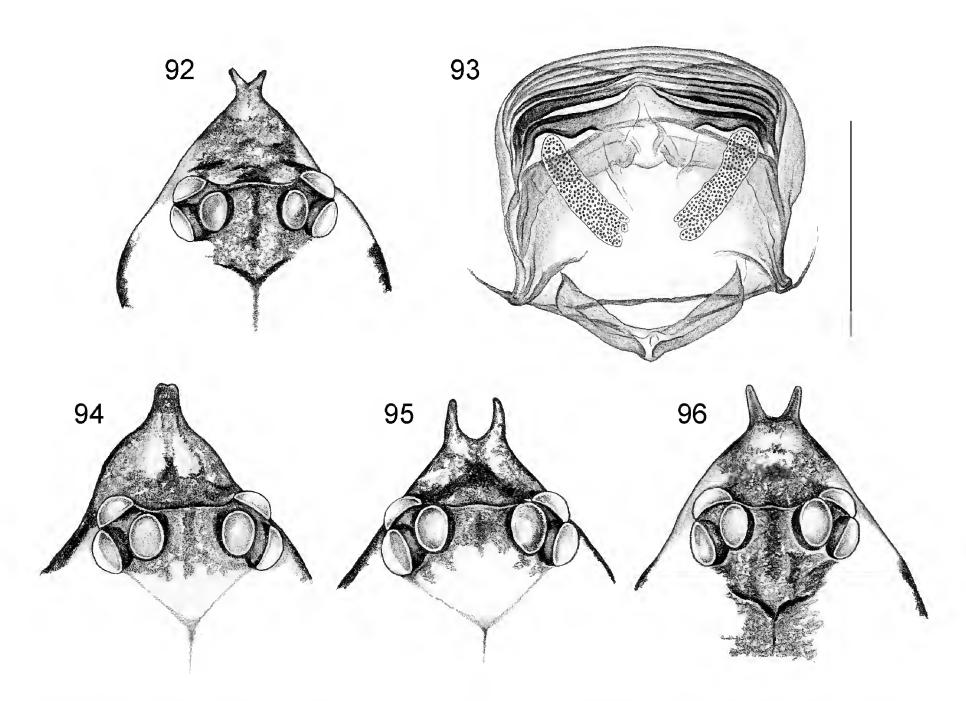
224319 part), same locality, fogging in primary rainforest, 4-12 Nov. 2006 and 26 Sep.-4 Oct. 2007 (D. De Bakker, J.P. Michiels). 1 \mathrew in MRAC (219999), same locality, beating along trail near guest house, 14 Nov. 2006 (D. De Bakker, J.P. Michiels). 4 \mathrew \m

Description

Male (holotype)

Measurements. Total body length 2.1, carapace width 0.7. Leg 1: 4.7 + 0.3 + 4.8 + 7.9, tarsus missing, tibia 2: 2.7, tibia 3: 1.8, tibia 4: 2.7; tibia 1 L/d: 68. Distance PME-PME 135 μ m, diameter PME 80 μ m, distance PME-ALE 25 μ m, AME absent.

COLOR. Carapace pale ochre-yellow with narrow black margins and indistinct median line (Fig. 84), ocular area and clypeus with dark pattern, sternum whitish (Fig. 89), legs ochre-yellow, with indistinct



Figs 92-96. *Spermophora abibae* sp. nov. and putative close relatives. **92-93**. *Spermophora abibae* sp. nov. \circlearrowleft ocular area and clypeus, dorsal view (92) and cleared \supsetneq genitalia, dorsal view (93). **94-96**. \circlearrowleft ocular area and clypeus, dorsal view of *S. dieke* Huber, 2009 (94) and of *S. akwamu* Huber & Kwapong, 2013 (95: Ghana; 96: Gabon). Scale line: 93 = 0.3 mm.

dark rings subdistally on femora and tibiae, abdomen mostly pale gray, with distinct dark pattern dorsally and laterally, ventrally only pair of marks at booklung covers.

Body. Habitus as in Fig. 84; ocular area slightly elevated, each triad on low hump; carapace without median furrow (only dark line); clypeus with distinctive bifid process, about 120 μm long (Figs 85, 92); sternum wider than long (0.52/0.44), unmodified. Chelicerae as in *S. dieke* and *S. akwamu* (cf. fig. 89 in Huber & Kwapong 2013), with pair of weakly sclerotized processes proximally, distally without modification, without stridulatory ridges.

Palps. As in Figs 86-88, apparently indistinguishable from *S. dieke* and *S. akwamu*; coxa with indistinct ventral process, trochanter with long ventral and shorter retrolateral apophyses, procursus with ventral sclerotized flap, prolatero-dorsal hinged process and thin distal flagellum; bulb with hooked apophysis and long weakly sclerotized embolus.

LEGS. Without spines and curved hairs, few vertical hairs; retrolateral trichobothrium on tibia 1 at 10%; prolateral trichobothrium absent on tibia 1, present on other tibiae.

Variation. Tibia 1 in 30 other males: 4.8-5.9 (mean 5.3), in one exceptional but otherwise indistinguishable male: 3.0.

Female

In general similar to male but sternum in most females dark brown or black (Fig. 90), clypeus unmodified, carapace with small median process acting against indistinct sclerotized plate above pedicel. Tibia 1 in 30 females: 3.7-4.9 (mean 4.3). Epigynum simple rectangular plate, anteriorly with dark transversal internal structure, without pockets (Fig. 91); internal genitalia as in Fig. 93.

Relationships

Judging from the very similar morphology (including the most probably derived median process on the female carapace), this species is probably closely related to *S. dieke* from Guinea and the Guineo-Congolian *S. akwamu*.

Distribution

Known from type locality in Congo D.R. only (Fig. 34).

Spermophora awalai Huber, sp. nov.

<u>urn:lsid:zoobank.org:act:0F2AB0B0-5E64-4F4E-9C10-D62EBE8BEAFA</u> Figs 5-6, 34, 97-107

Diagnosis

Easily distinguished from most congeners by procursus (Figs 102-103; strongly bent towards ventral; with large hinged ventral process); from similar (East African) species (*S. morogoro* Huber, 2003; *S. usambara* Huber, 2003; *S. masisiwe* Huber, 2003; cf. figs 222, 239, 245 in Huber 2003b) by long apophyses on male chelicerae (Figs 104-105); from these and other congeners also by distinctive pair of round structures with concentric rings in female internal genitalia (Figs 98, 106).

Etymology

Named for the Cameroonian writer Alexandre Biyidi Awala (1932-2001), known as Mongo Beti.

Type material

Holotype ♂, in ZFMK (Ar 11953).

Type locality

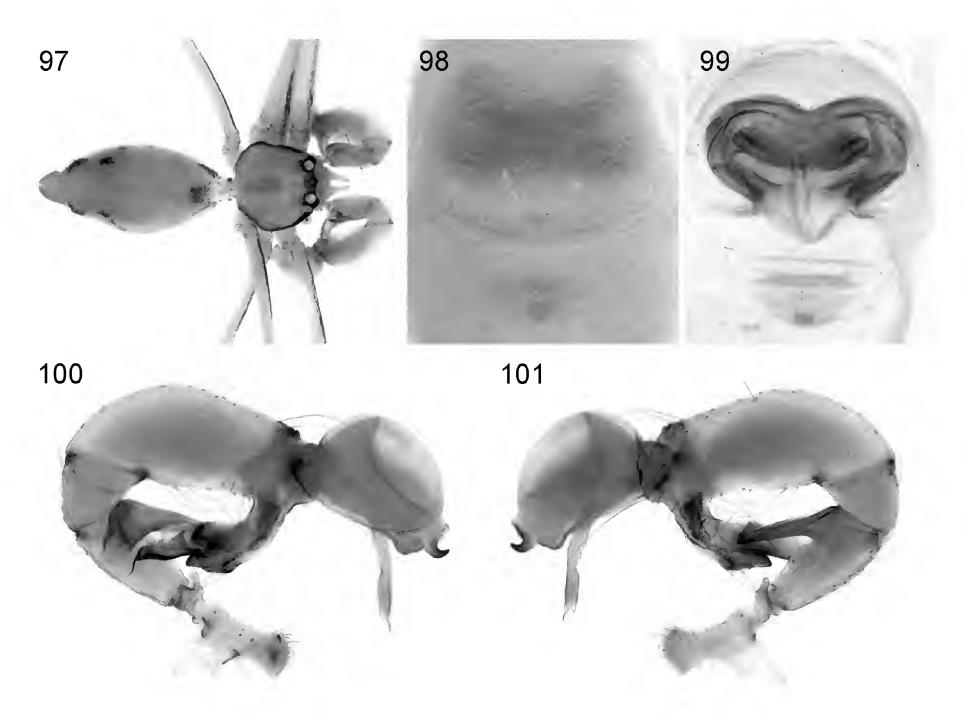
CAMEROON, Southwest Region, near Dschang, Attuleh, 'site 2' (5°27.9'N, 9°56.5'E), 1800 m a.s.l., underside of leaves, 20 Apr. 2009 (B.A. & J.C. Huber).

Other material examined

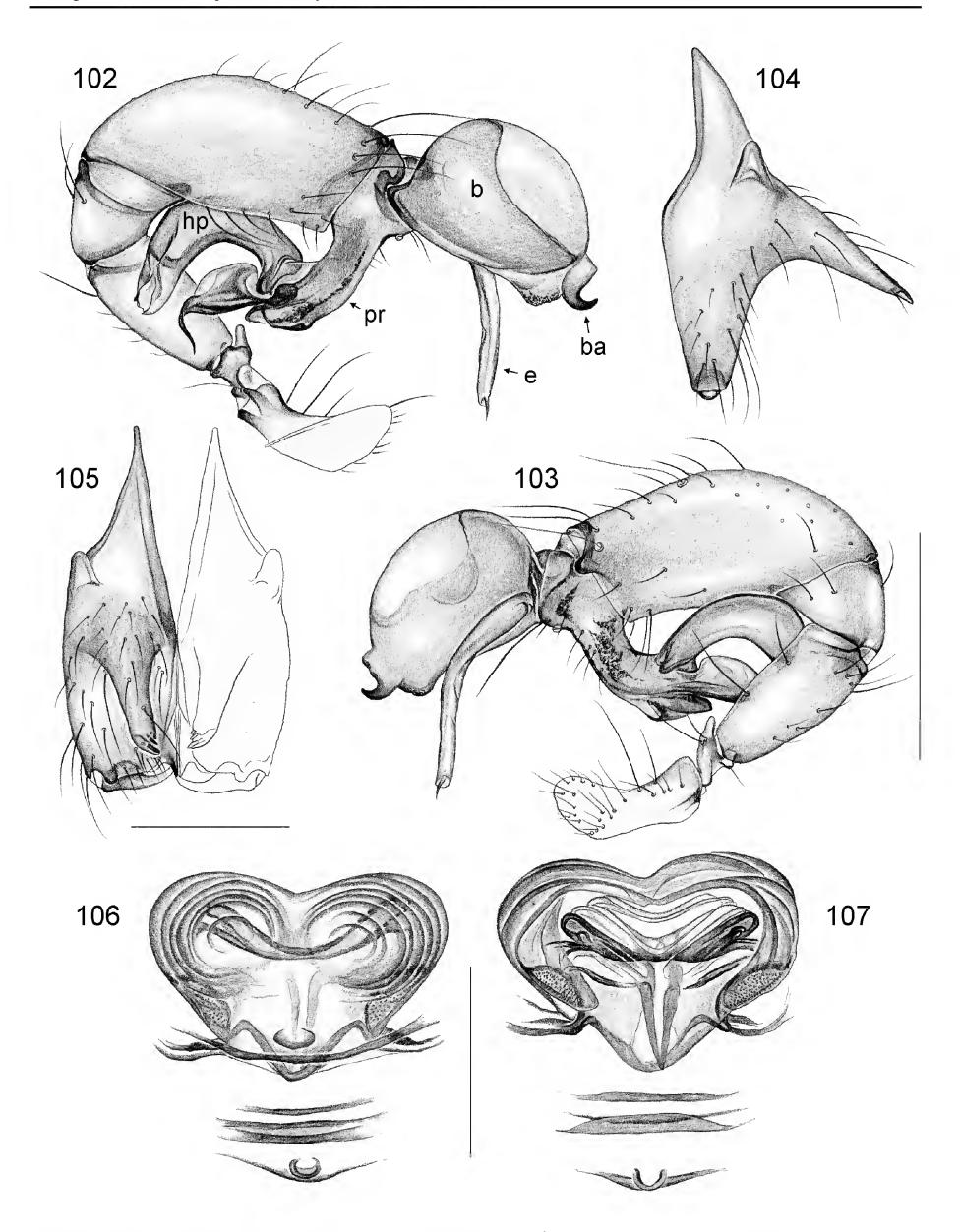
Description

Male (holotype)

Measurements. Total body length 2.2, carapace width 0.8. Leg 1: 21.5 (5.2 + 0.3 + 5.2 + 7.9 + 2.9), tibia 2: 3.3, tibia 3: 2.0, tibia 4: 2.9; tibia 1 L/d: 73. Distance PME-PME 220 μ m, diameter PME 95 μ m, distance PME-ALE 35 μ m, AME absent.



Figs 97-101. Spermophora awalai sp. nov. 97. ♂, dorsal view. 98. ♀ epigynum, ventral view. 99. Cleared ♀ genitalia, dorsal view. 100-101. Left ♂ palp, prolateral and retrolateral views.



Figs 102-107. *Spermophora awalai* sp. nov. **102-103**. Left \bigcirc palp, prolateral and retrolateral views. **104-105**. \bigcirc chelicerae, lateral and frontal views. **106-107**. Cleared \bigcirc genitalia, ventral and dorsal views. b = bulb; ba = bulbal apophysis; e = embolus; hp = hinged process; pr = procursus. Scale lines: 102-103 = 0.5 mm; 104-105 = 0.2 mm; 106-107 = 0.3 mm.

Color. Carapace ochre-yellow with black lateral margins and light brown median mark (Fig. 97), ocular area with dark band between eye triads, clypeus with a few black spots, sternum whitish, legs ochre-yellow, with black spots on femora proximally, black patellae and tibia-metatarsus joints, dark rings subdistally on femora and tibiae (unusually far from tip), abdomen ochre gray with black marks dorsally and laterally.

Body. Habitus as in Fig. 97; ocular area barely elevated, each triad on low hump; carapace without median furrow; clypeus unmodified; sternum wider than long (0.58/0.42), unmodified. Chelicerae as in Figs 104-105, with pair of proximal lateral apophyses and long frontal apophyses provided with three modified hairs each at tips, without stridulatory ridges.

Palps. As in Figs 100-103, coxa unmodified, trochanter with short retrolatero-ventral apophysis, femur unmodified, procursus strongly bent towards ventral, with complex tip and large ventral hinged process; bulb with long, weakly sclerotized embolus and short, hooked apophysis.

LEGS. Without spines and curved hairs, few vertical hairs; retrolateral trichobothrium on tibia 1 at 8%; prolateral trichobothrium absent on tibia 1, present on other tibiae. Tarsus 1 with >20 pseudosegments, only distally fairly distinct.

Variation. Tibia 1 in 12 other males: 4.1-5.5 (mean 4.9). Some specimens with additional indistinct dark ring on each femur and tibia.

Female

In general similar to male; tibia 1 in 10 females: 3.6-4.6 (mean 4.2). Epigynum light brown area with pair of distinctive internal structures with concentric rings visible through cuticle (Figs 98, 106), with median pocket between epigynum and spinnerets; internal genitalia as in Figs 99 and 107.

Distribution

Known from several localities in Southwest and Centre Regions, Cameroon (Fig. 34).

Discussion

Distribution patterns

Except for some synanthropic and widespread species, the large majority of Central African Pholcidae are highly endemic. Considering just the two neighboring countries that were relatively well sampled (Cameroon and Gabon), the number of endemic species is considerably higher than the number of non-endemic species in each country. In Cameroon, 21 of the 35 currently recorded species (i.e. 60%) are not known from any other country, not even neighboring Gabon. The numbers for Gabon are almost identical (21 of 33 species endemic, i.e. 64%). Diversity and endemism are especially high in the genus *Smeringopina*, with 13 of the 14 Cameroonian species and all 18 Gabonese species known from only Cameroon and Gabon, respectively.

Most of the species listed above (62, i.e. 84% if the five synanthropic species are discounted from the total number of 79) are endemics of the Lower Guinean subregion of the Guineo-Congolian center of endemism. Similar percentages of endemics have been found in West Africa (68%; value differs from that in Huber & Kwapong 2013 because *Anansus atewa* is no longer considered an endemic of West Africa) and East Africa (84%; calculated from data in Huber & Warui 2012). This is reflected in the fact that few species (again, discounting synanthropics) are shared among these regions. Only three species occur in all three regions: *Pehrforsskalia conopyga* Deeleman-Reinhold & van Harten, 2001; *Quamtana kitahurira* Huber, 2003 and *Spermophora kyambura* Huber & Warui, 2012. Three additional species are shared between Upper and Lower Guinea (i.e., West and Central Africa): *Smeringopina fon* Huber,

2013; *Smeringopus cylindrogaster* (Simon, 1907); and *Spermophora akwamu* Huber & Kwapong, 2013 (a fourth, *Pholcus kakum* Huber, 2009, is likely to occur in Lower Guinea but has not yet been documented from there). Two Central African species range deeply into Congolia (*Pholcus baka* Huber, 2011 and *Smeringopus lesserti* Kraus, 1957), the latter reaching the western limits of East Africa (Huber 2011b, 2012).

A comparison of the genera represented by endemic species in each of the three regions studied (Fig. 108) reveals significant differences between East and Central Africa but smaller differences between Central and West Africa. This is mainly due to the species-rich genera *Buitinga* Huber, 2003 and *Smeringopina* Kraus, 1957, the first being endemic to East Africa, the second to West and Central Africa. By contrast, the percentages of endemic species of *Pholcus* Walckenaer, 1805 are relatively homogeneous across tropical Africa. This is noteworthy because *Pholcus* has no endemic species in southern Africa or on Madagascar (Huber 2011b).

Diversity patterns

From a simple comparison of total species numbers, pholcid diversities in Central and East Africa appear to be similar (79 and 86 species respectively), while West Africa seems to be significantly less diverse (38 species; Fig. 109). This is not simply a result of different surface areas considered in each case. The land area of West Africa considered in Huber & Kwapong (2013; ~2.3 x 10⁶ km²) was actually larger than that of East Africa in Huber & Warui (2012; ~2.1 x 10⁶ km²) and that of Central Africa considered herein (~1.6 x 10⁶ km²). On the other hand, there is some evidence that the total numbers are biased against Central Africa because it seems most poorly collected. Figure 110 shows species numbers collected by the first author during expeditions of very similar durations and with very similar collecting efforts (West Africa: Guinea, Ghana; Central Africa: Cameroon, Gabon; East Africa: Kenya, Uganda). The much higher number of new species found in Central Africa suggests that this region is in fact undersampled relative to West and East Africa. Recent evidence on forest-dwelling insects suggests that the diversity is higher in Central Africa (and in particular in Cameroon) than in any other part of Africa (P. Le Gall, unpublished).

Diversity is also high at individual localities (with 'locality' roughly defined as the area that can be covered in one day by walking). Few localities worldwide are known to contain more than 10 species of

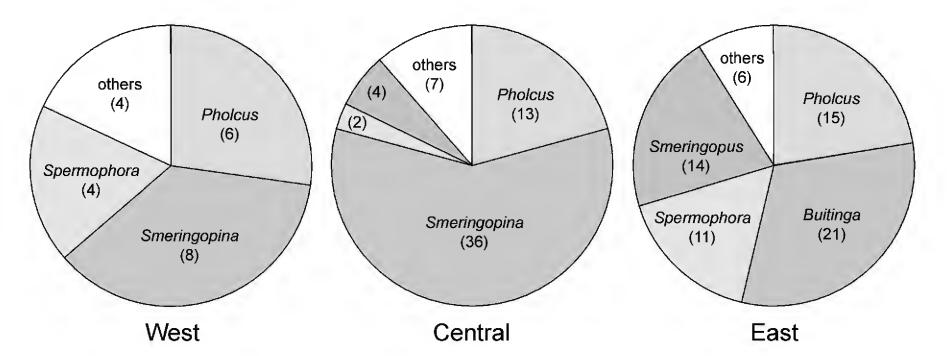
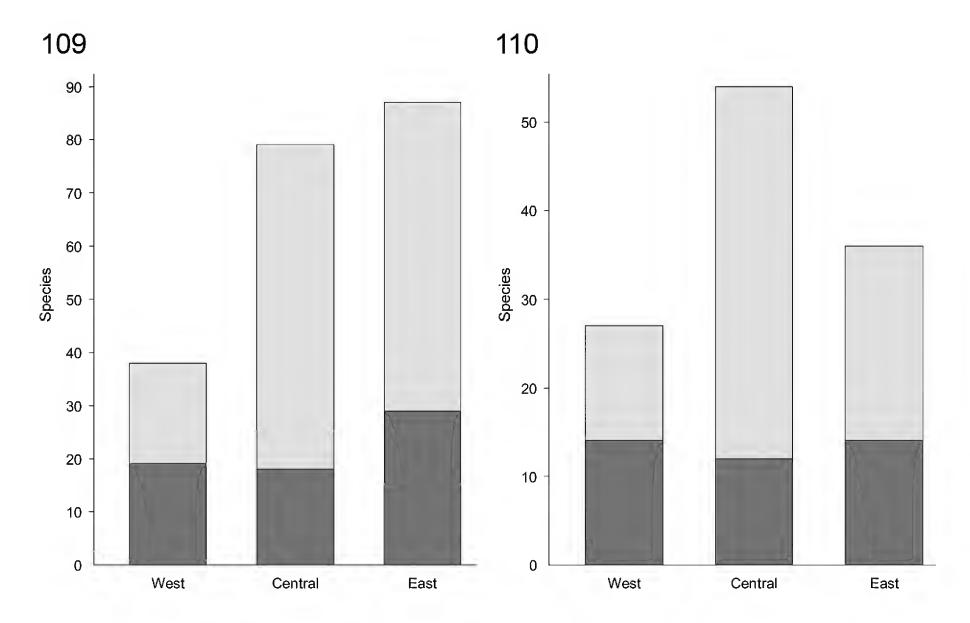


Fig. 108. Pholcid genera represented by endemic species (total numbers of endemic species in parentheses) in each region (West, Central, and East Africa), illustrating the distinctness of East Africa as compared to West and Central Africa. Data on East and West Africa from Huber & Warui (2012) and Huber & Kwapong (2013), respectively. White: genera that are not represented by more than four endemic species in any of the three regions.

Pholcidae each, most of them in Brazil's Atlantic Forest (Huber & Rheims 2011; B.A. Huber, unpublished data). Luki Forest Reserve in Central Africa is currently the most diverse locality in Africa for this family. Eleven of its 13 known species (the last of which is described herein), have been described within the last decade: *Anansus debakkeri* Huber, 2007; *Leptopholcus debakkeri* Huber, 2011; *L. tipula* (Simon, 1907); *Pholcus luki* Huber, 2011; *P. soukous* Huber, 2011; *Quamtana kitahurira* Huber, 2003; *Smeringopina kikongo* Huber, 2013; *S. luki* Huber, 2013; *Smeringopus luki* Huber, 2012; *S. mayombe* Huber, 2012; *Spermophora abibae* sp. nov.; *S. kyambura* Huber & Warui, 2012; *Nyikoa limbe* Huber, 2007. Since Luki Forest Reserve is probably among the best sampled African localities as far as spiders are concerned (as a result of two major campaigns by MRAC focusing on this single forest), it is likely that increased effort will reveal more such diverse localities in Central Africa. The fact that 10 species were collected at Mayebout (Gabon) within two days by only two collectors (one of them inexperienced) also points in that direction.

Outlook

Two recent expeditions to Cameroon and Gabon resulted in a total of 54 pholcid species, 42 of which (78%) were still undescribed a decade ago. All this material and most of the Central African material deposited in other major arachnological collections is now described. However, the high percentage of undescribed species collected during about seven weeks in the field also illustrates the relative ease with which new pholcid species can still be discovered in this region. This is probably a general pattern, as suggested by recent studies on arthropods in lowland forests in Cameroon (e.g., Larsen 2005) and on



Figs 109-110. Pholcid species from West, Central, and East Africa, divided into 'old' species (grey; described before 2003) and 'new' species (green; described since 2003). **109**. Current totals as listed in Huber & Warui (2012), Huber & Kwapong (2013) and herein, suggesting lower diversity in West than in Central and East Africa. **110**. Species collected by the first author during six expeditions (two to each region) of very similar durations and with very similar collecting efforts, suggesting that Central Africa may be more severely undersampled than West and East Africa.

the biodiversity of the Volcanic Line of Cameroon (Bergl *et al.* 2007). Species distribution patterns also suggest that even in areas relatively well sampled (western Cameroon, Gabon, western Congo D.R.), many undiscovered species may remain. Of the 79 Central African species listed above, 33 (42%) are known from their type localities only. Denser sampling across seasons and altitudes will probably reveal not only more realistic distribution patterns, but also additional new species even in these areas.

On the other hand, large areas within Lower Guinea remain almost entirely unexplored with respect to spiders and to Pholcidae in particular (Fig. 19). Congo Republic is among the few countries worldwide from which the first author has never seen even a single pholcid specimen. Only a single (doubtful) record seems to exist in the literature (a juvenile *Smeringopus pallidus* cited in Lawrence 1958). With large parts of its moist tropical forests remaining (over 60% as of 1992; Naughton-Treves & Weber 2001), Congo Republic is likely to have similar numbers of species as neighboring Gabon (currently 33). The same is true for large parts of southeastern Cameroon, eastern Nigeria, and southwestern Central African Republic (Fig. 19).

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